

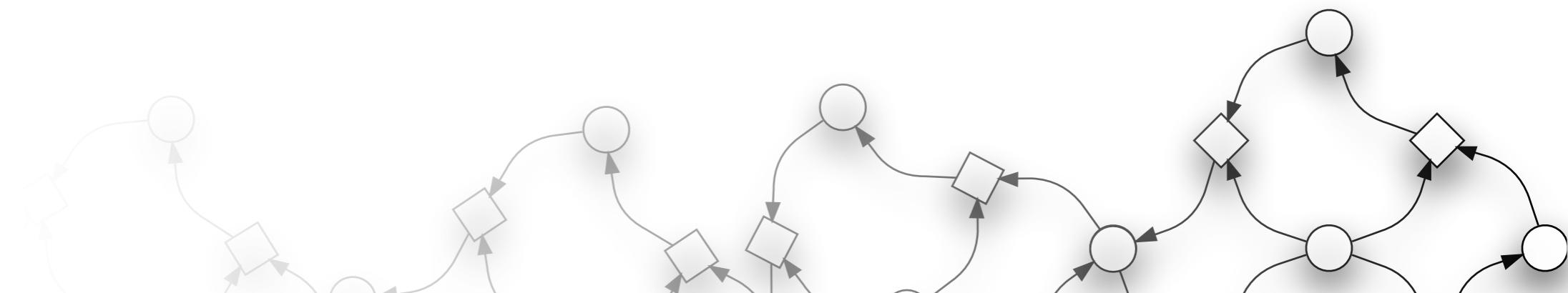
BioPPN 2015, Brussel/Belgium - 22nd June 2015

Spatial Modelling based on the BMK-Framework

Mary-Ann Blätke¹, Christian Rohr²

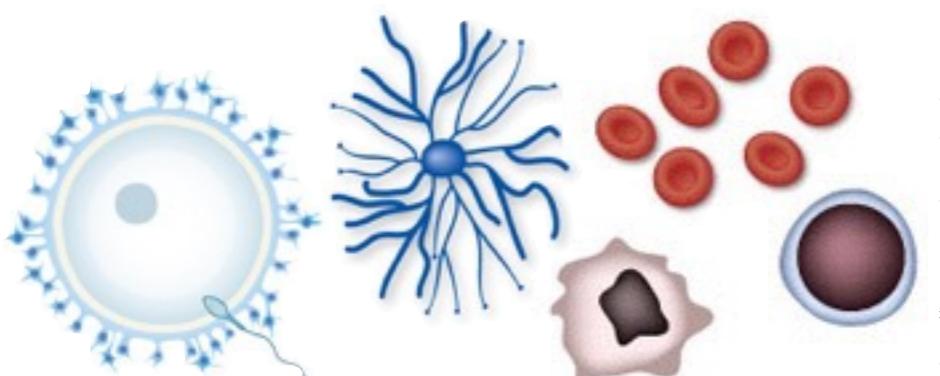
¹ Otto-von-Guericke-University Magdeburg, Germany

² Brandenburg University of Technology Cottbus, Germany



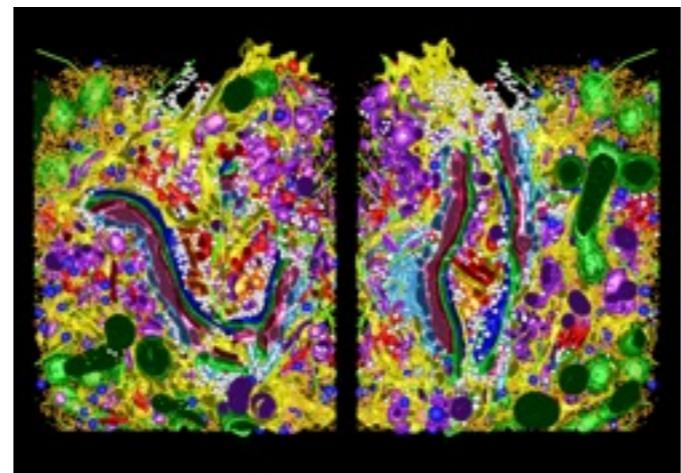
Background

- Systems Biology encouraged the modelling of bio-systems
 - detailed mechanistic & kinetic descriptions
 - ignore spatial aspects
- But space matters:



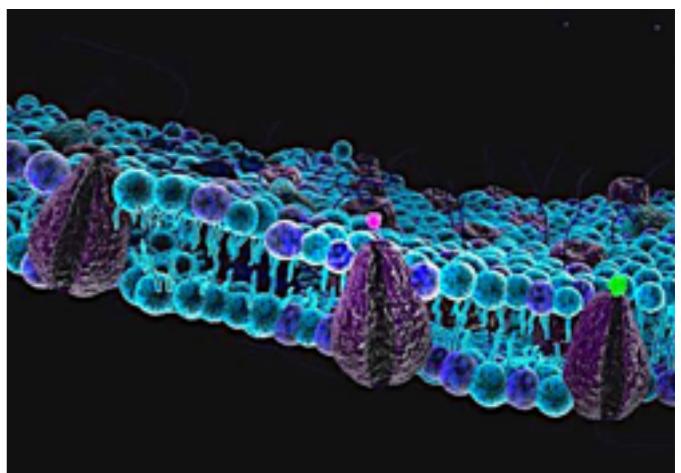
<http://learn.genetics.utah.edu>

cell size & shape



<http://learn.genetics.utah.edu>

compartmentalisation,
pools & organisation



<http://learn.genetics.utah.edu>

membrane composition

- Integration of spatial aspects in existing models is not straightforward
 - ➔ models have to be rebuild

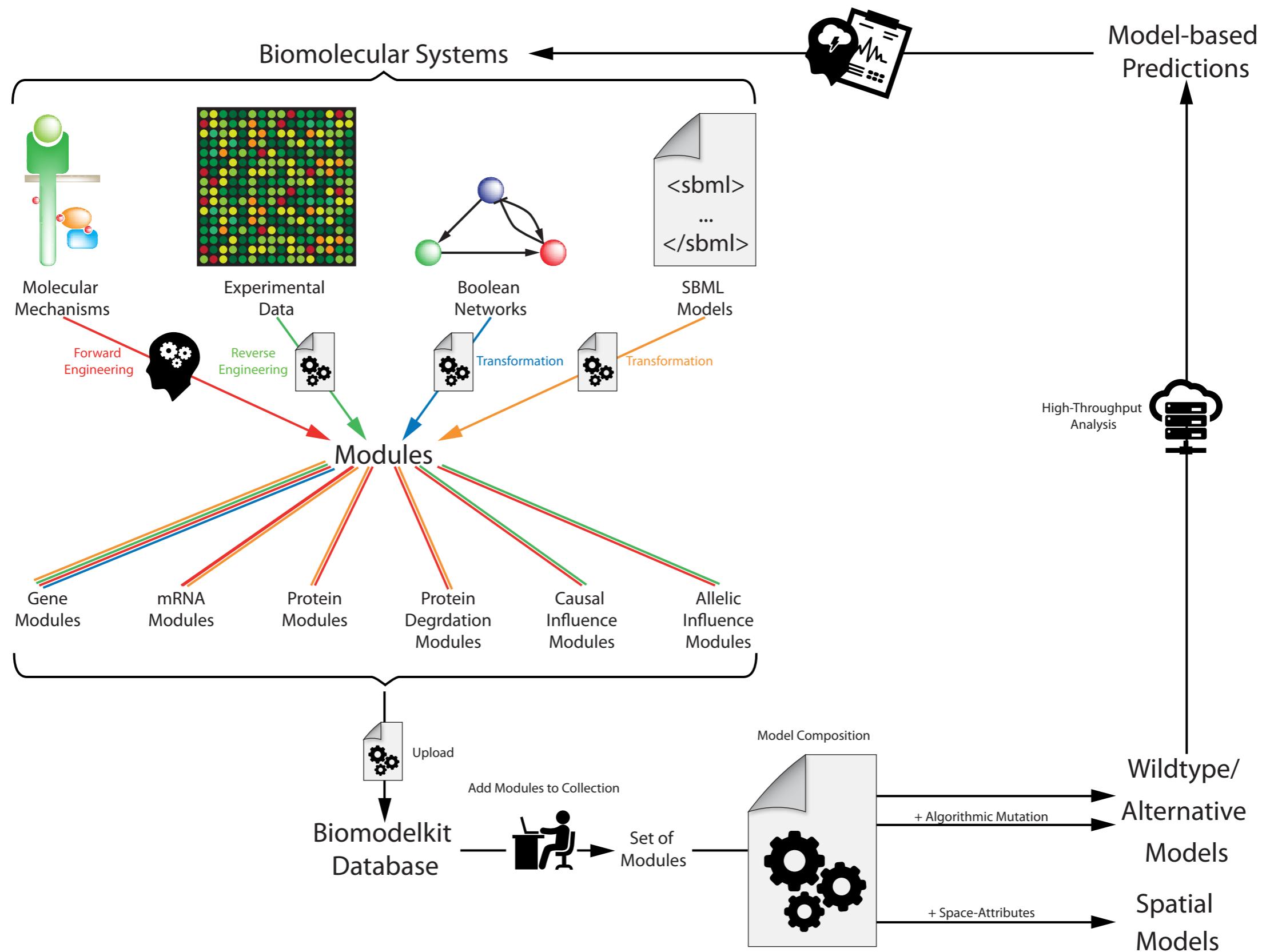
Our Idea

- Extend models with spatial information to keep track of position and movement
- Biomolecule-centred modularisation of models

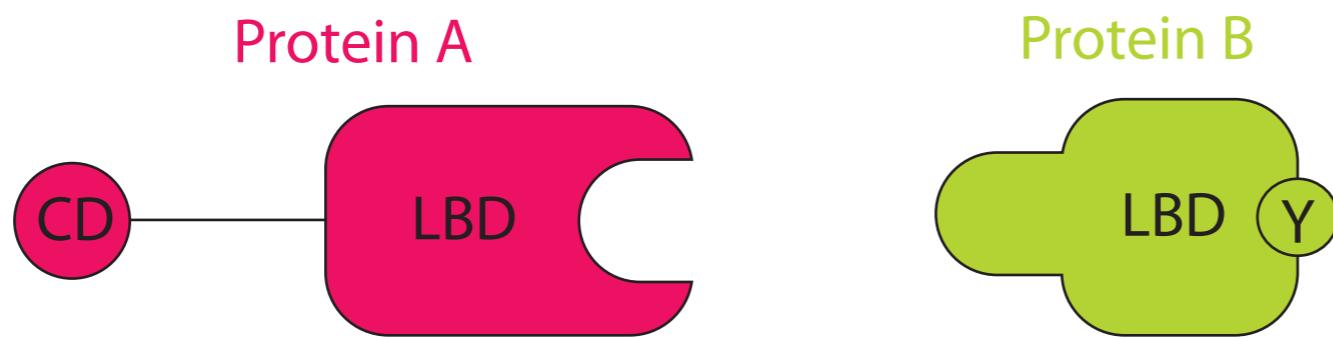
→ **Biomodelkit Framework**

A module describes the functionality and interaction of a biomolecule in the form of a Petri net

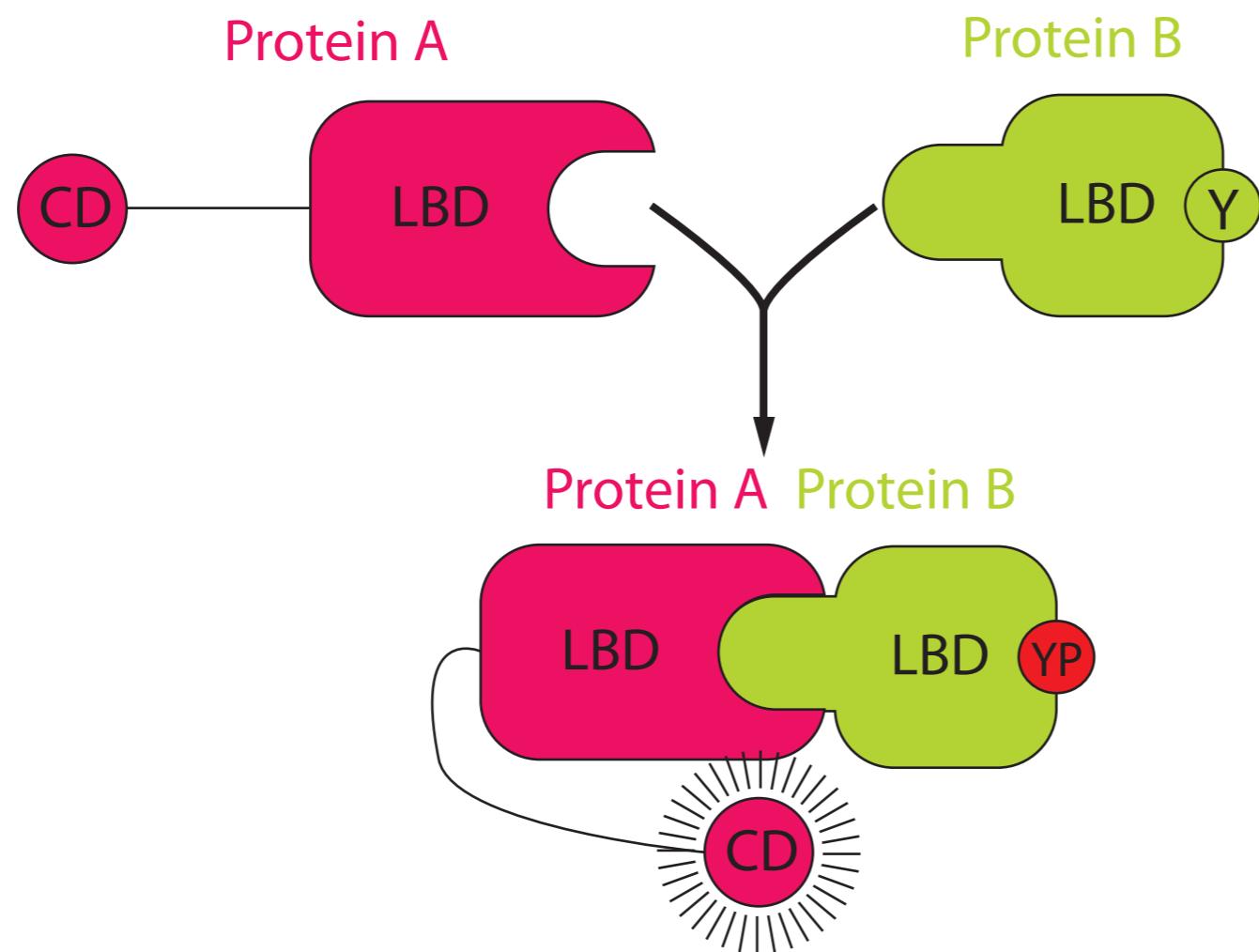
BMK-Framework



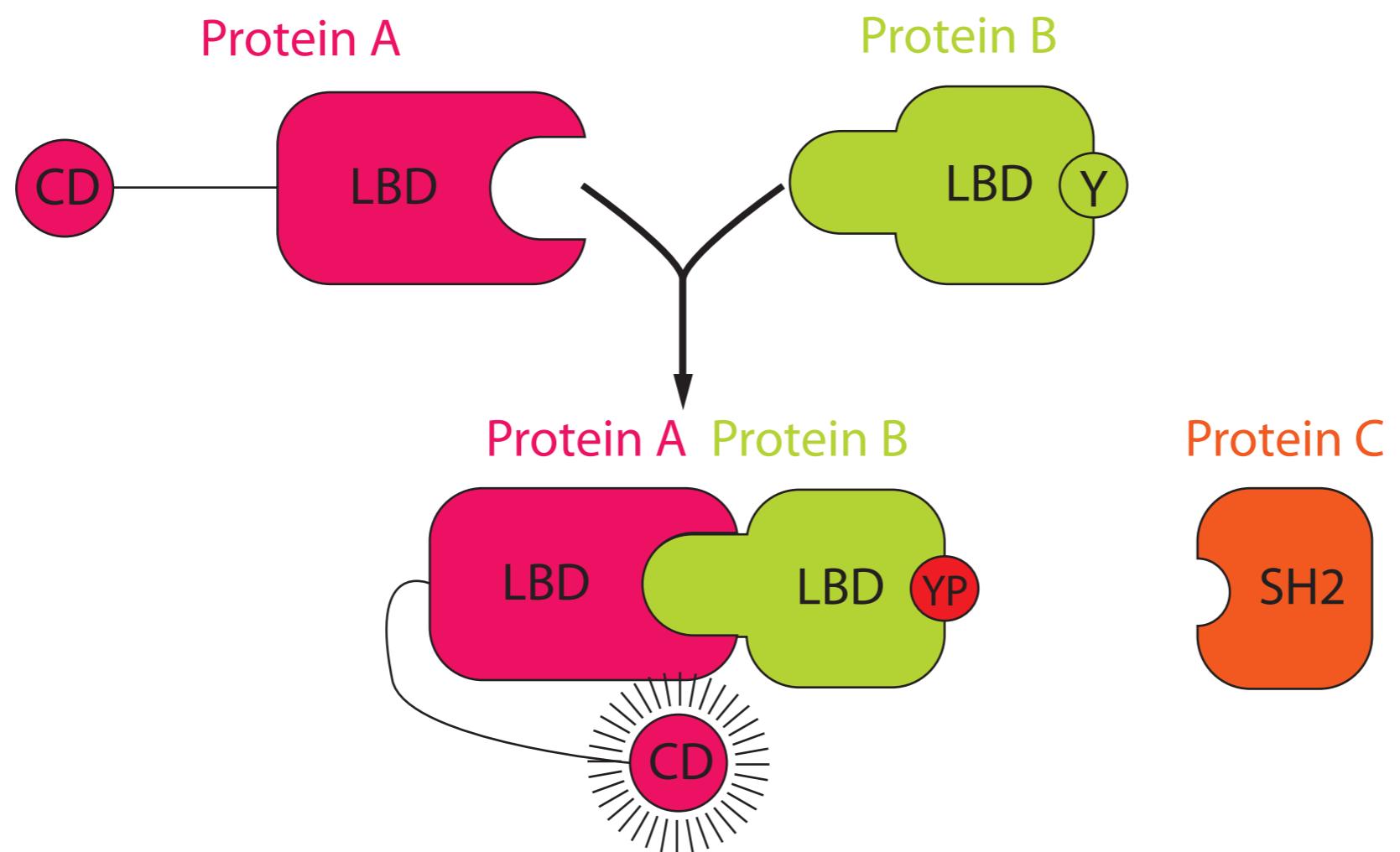
Running Example



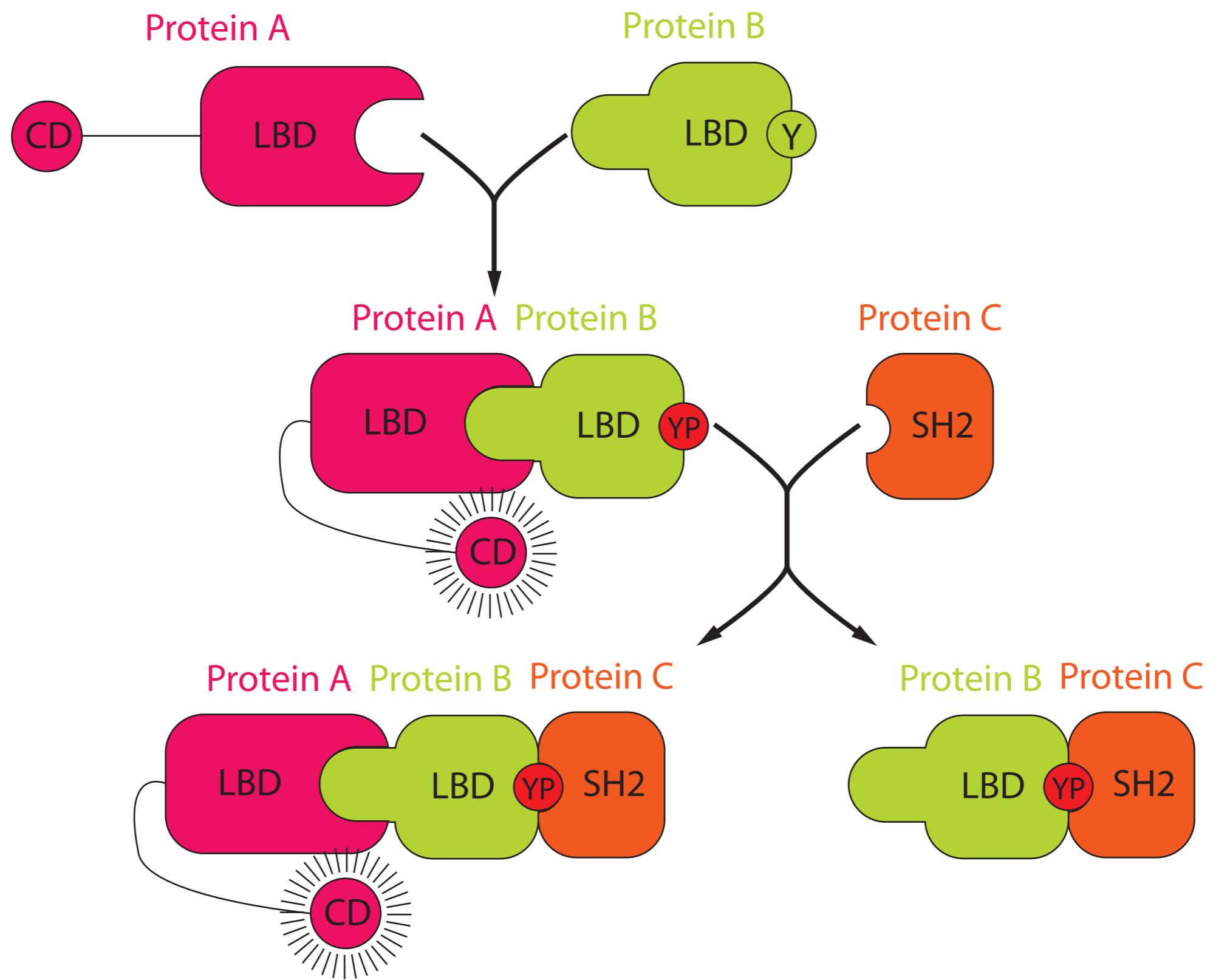
Running Example



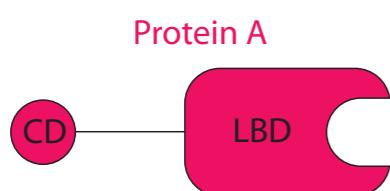
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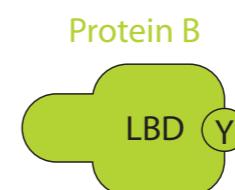
Running Example



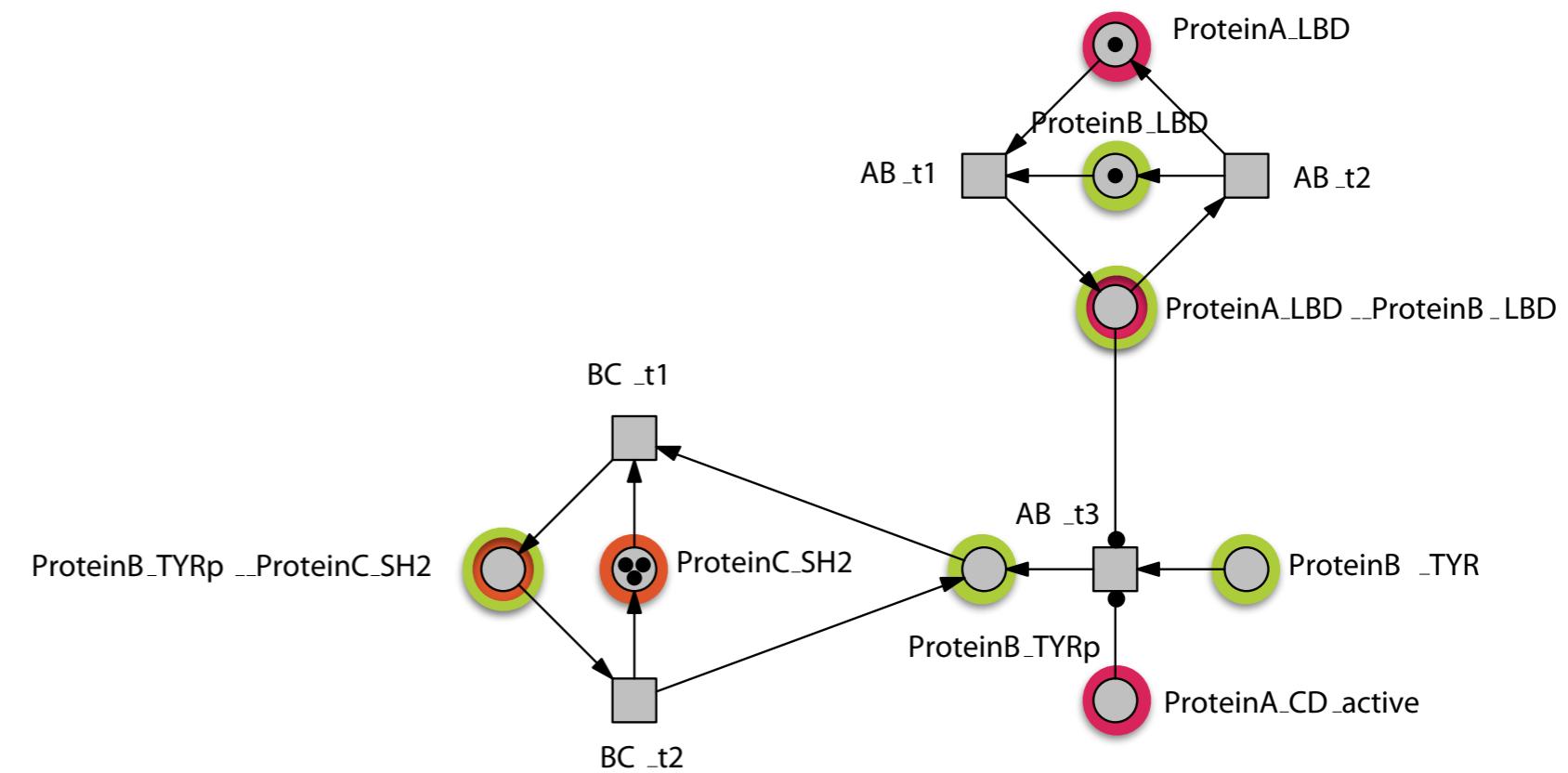
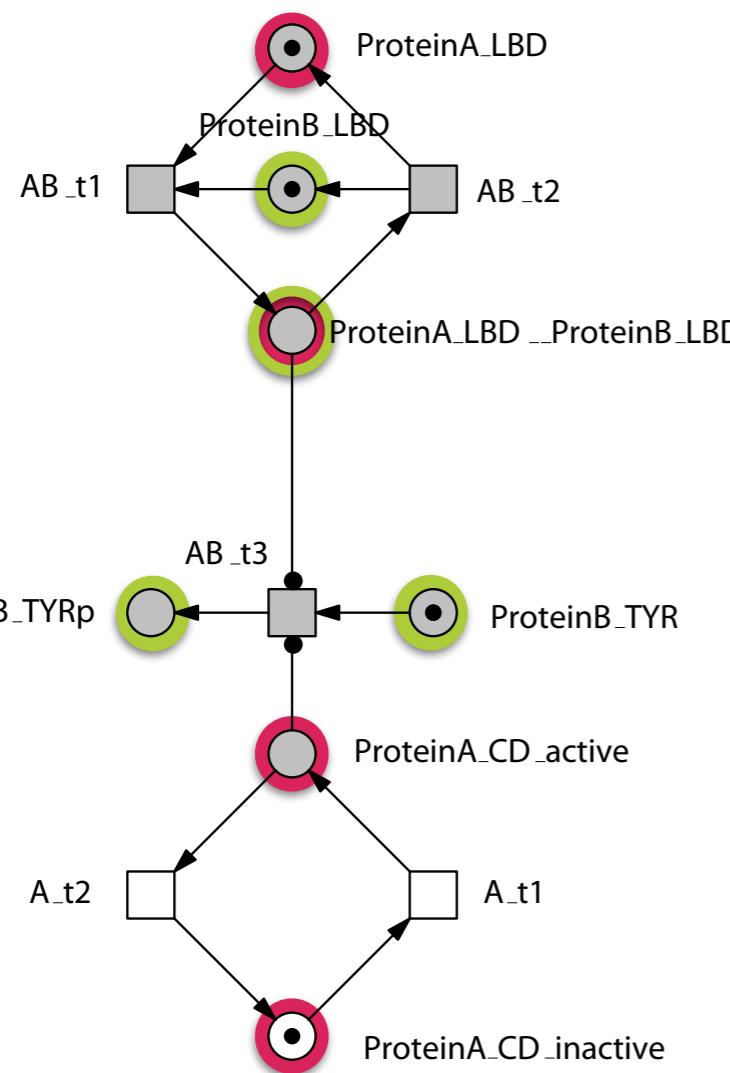
Modules of the Running Example



Module of Protein A

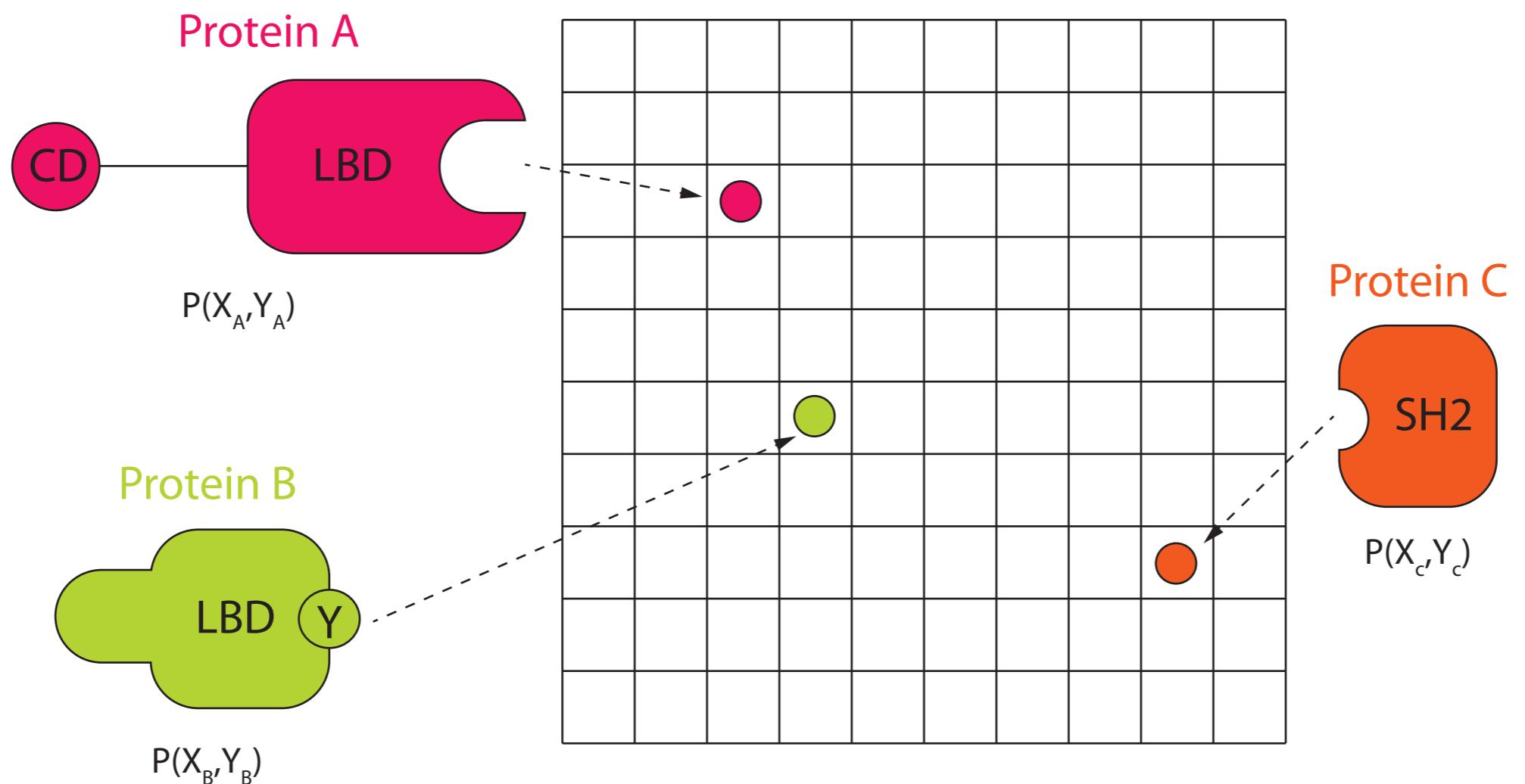


Module of Protein B



Step 1: Explicit Encoding of Local Positions

- Define a grid for each component (1D,2D or 3D)
 - e.g. 2D Grid: $x\text{DimA} = 10$, $y\text{DimA} = 10$
- Set a local position for each component
 - e.g. A(3,8), B(4,5), C(9,3)

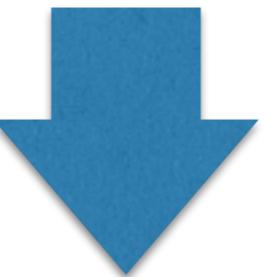


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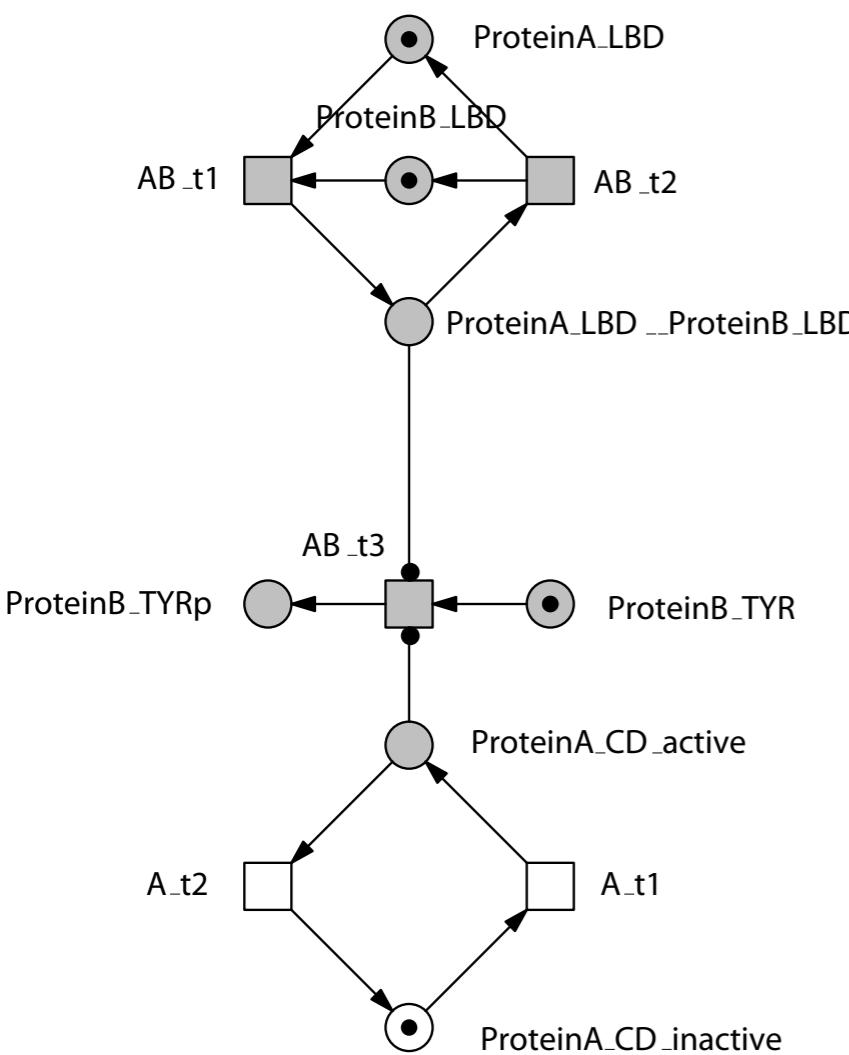
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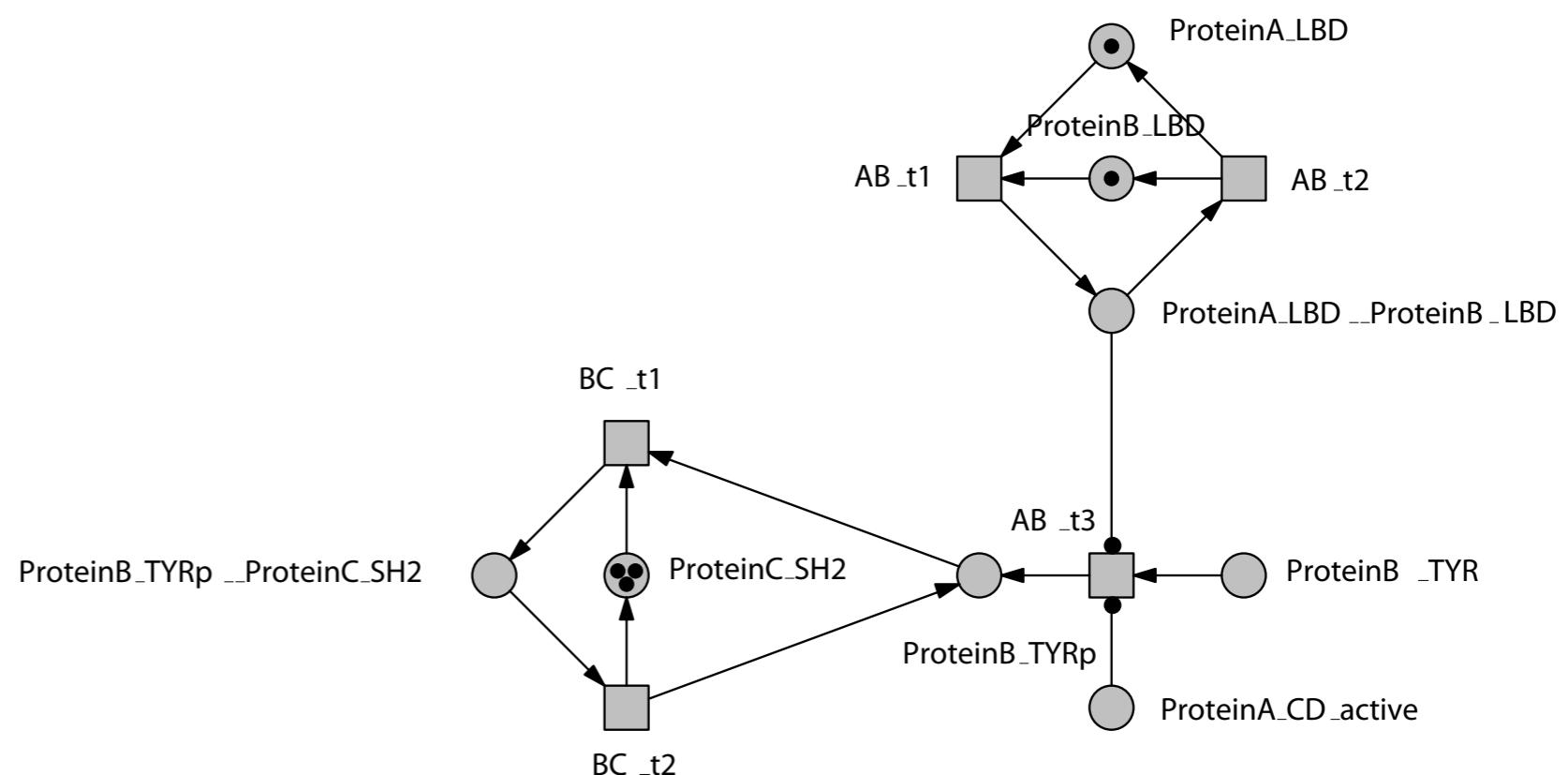
Add a place for each component and axis

Step 1: Explicit Encoding of Local Positions

Module of Protein A



Module of Protein B



XY-Position of Protein A



ProteinA_X

ProteinA_Y

XY-Position of Protein B



ProteinB_X

ProteinB_Y

XY-Position of Protein C

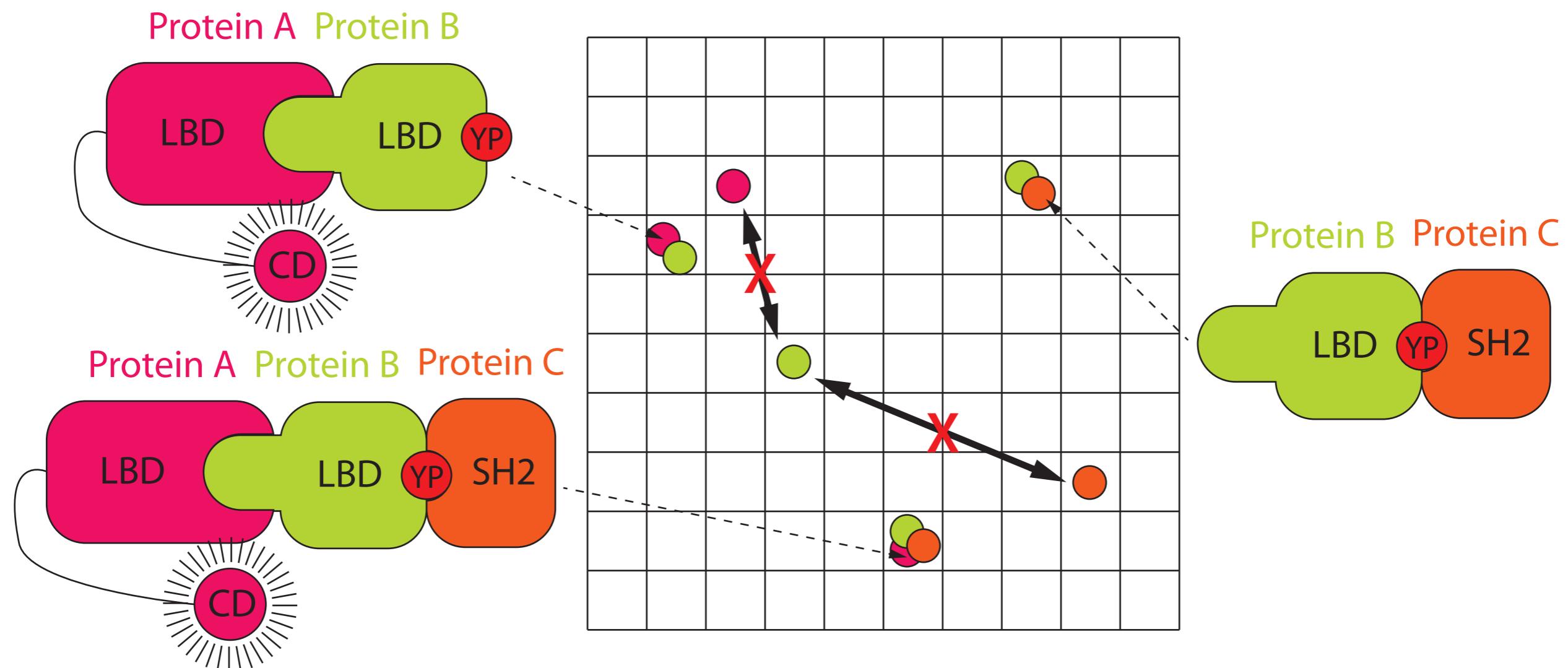


ProteinC_X

ProteinC_Y

Step 2: Local Restriction of Interactions

- Components can only interact if they fulfil a defined neighbourhood relation
- Define a neighbourhood relation
 - e.g. local positions of components must be identical

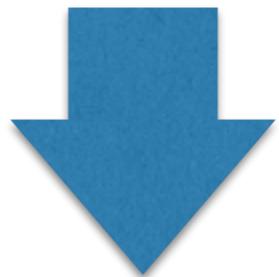


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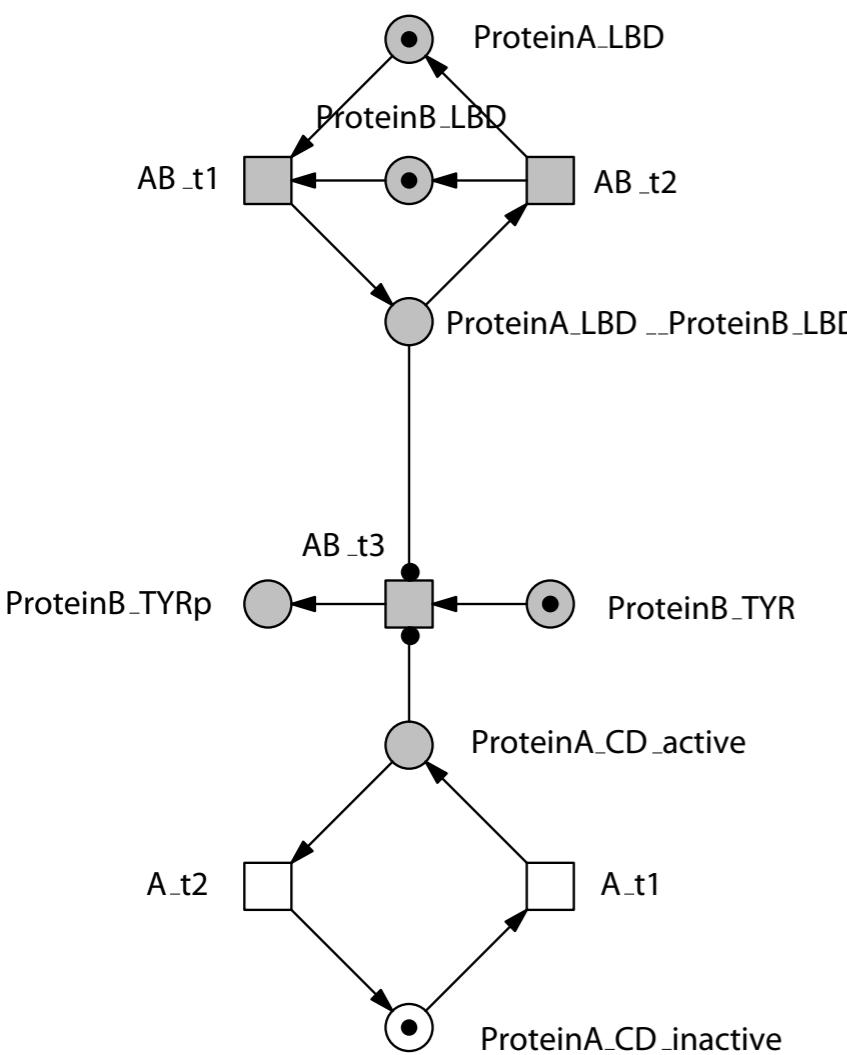


For each transition representing an interaction:

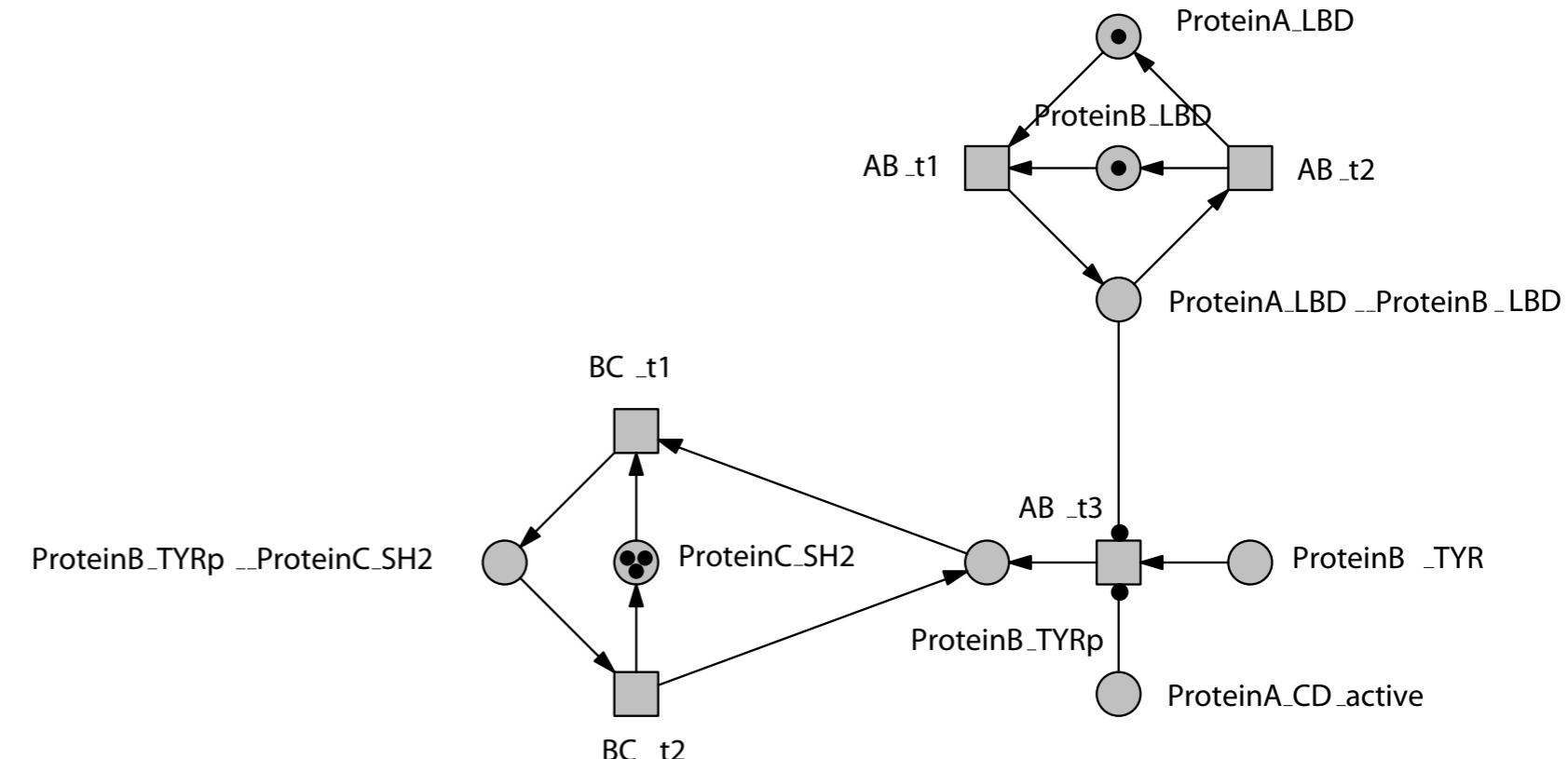
- add coordinate places of interacting components
- multiply firing-rate with a boolean expression evaluating the distance between the interacting components

Step 2: Local Restriction of Interactions

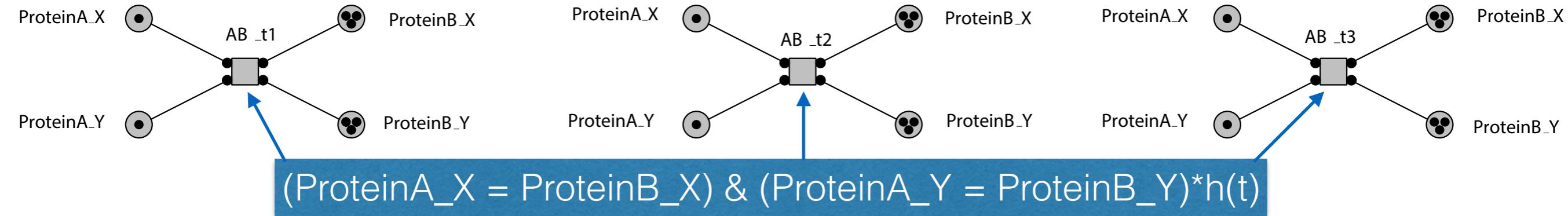
Module of Protein A



Module of Protein B

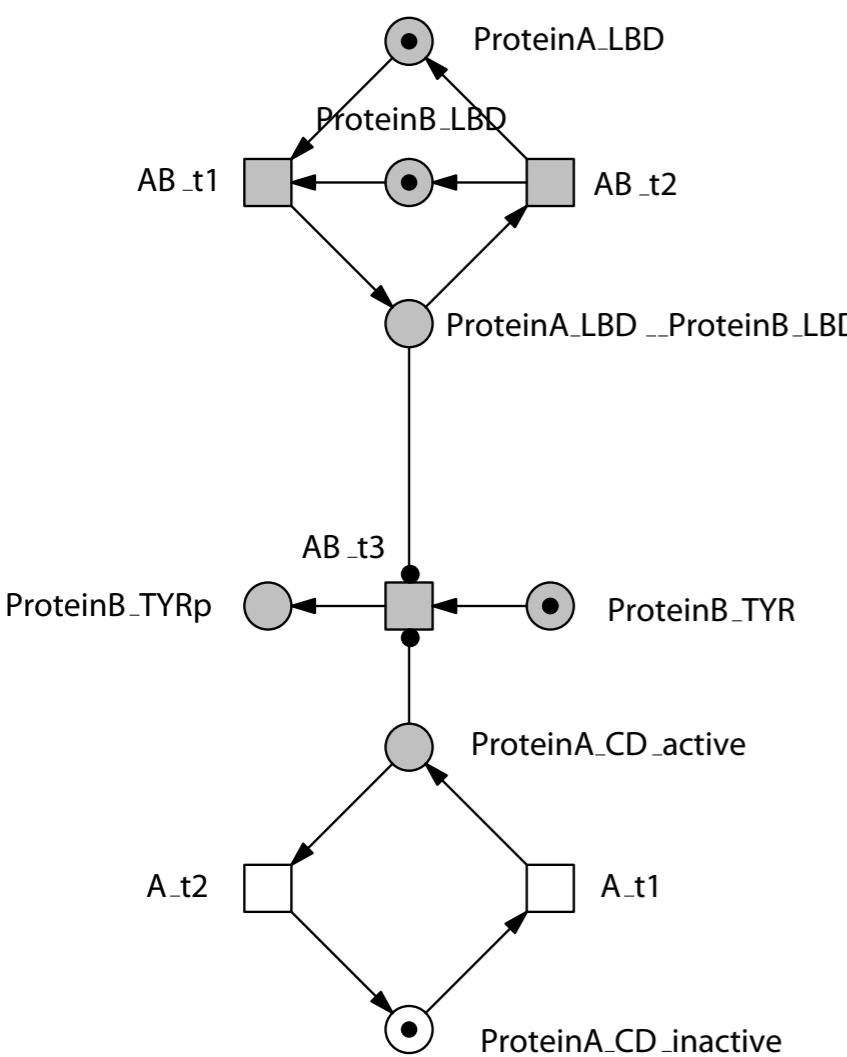


Restricted Interaction - Only if ProteinA (X,Y) = ProteinB (X,Y)

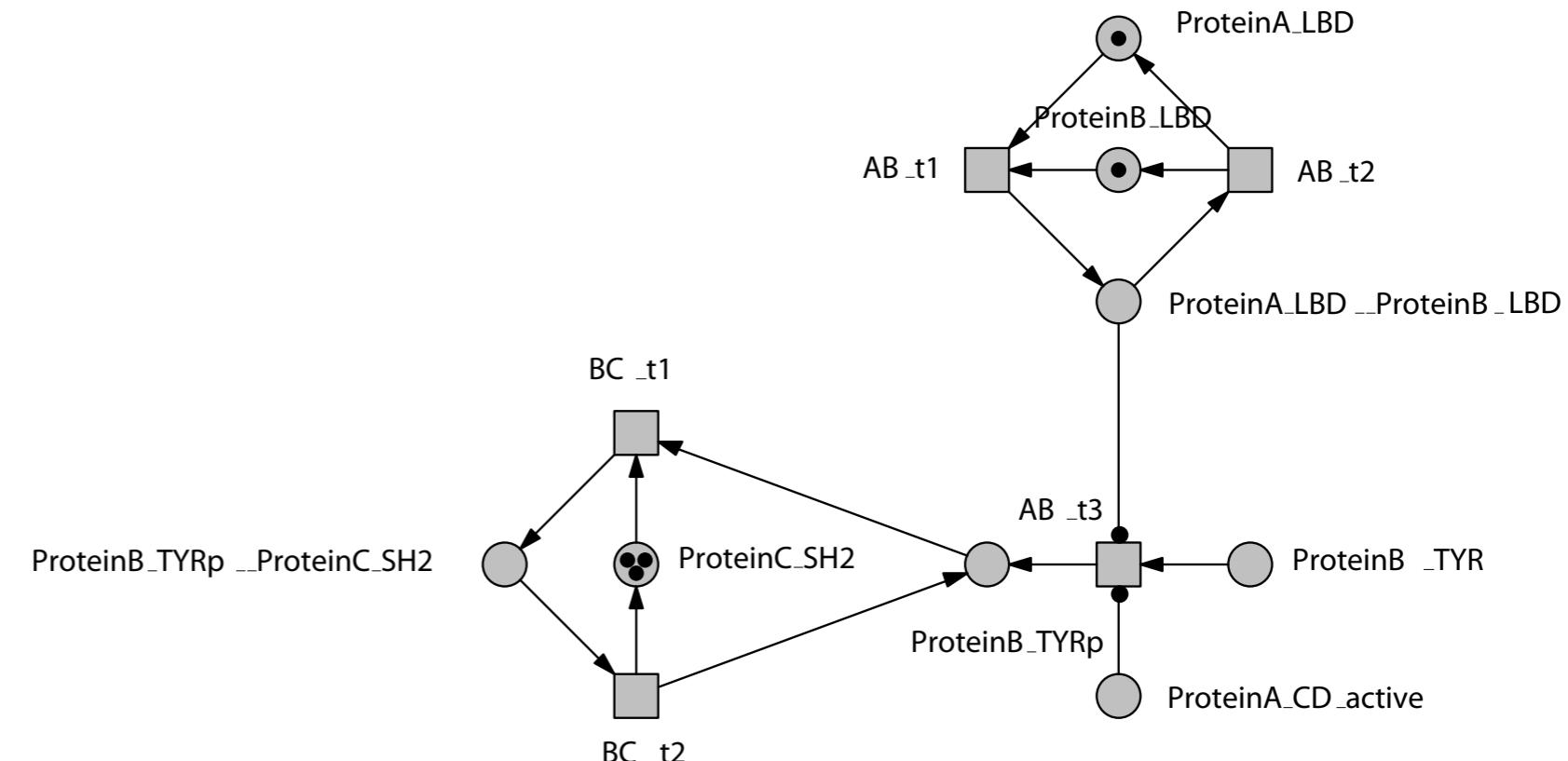


Step 2: Local Restriction of Interactions

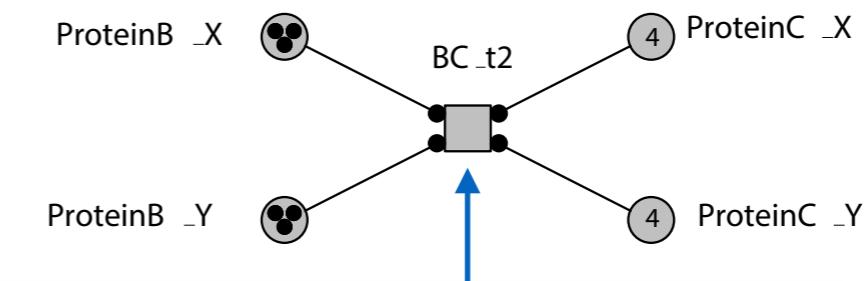
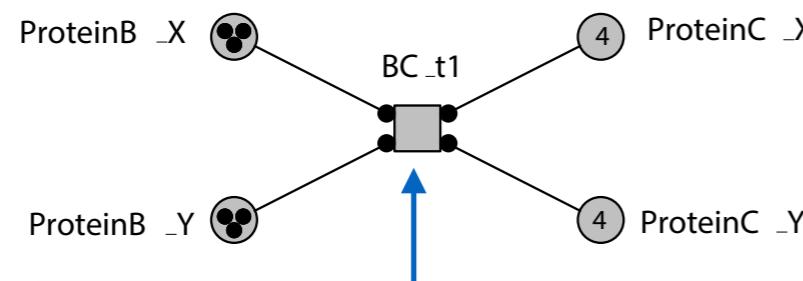
Module of Protein A



Module of Protein B



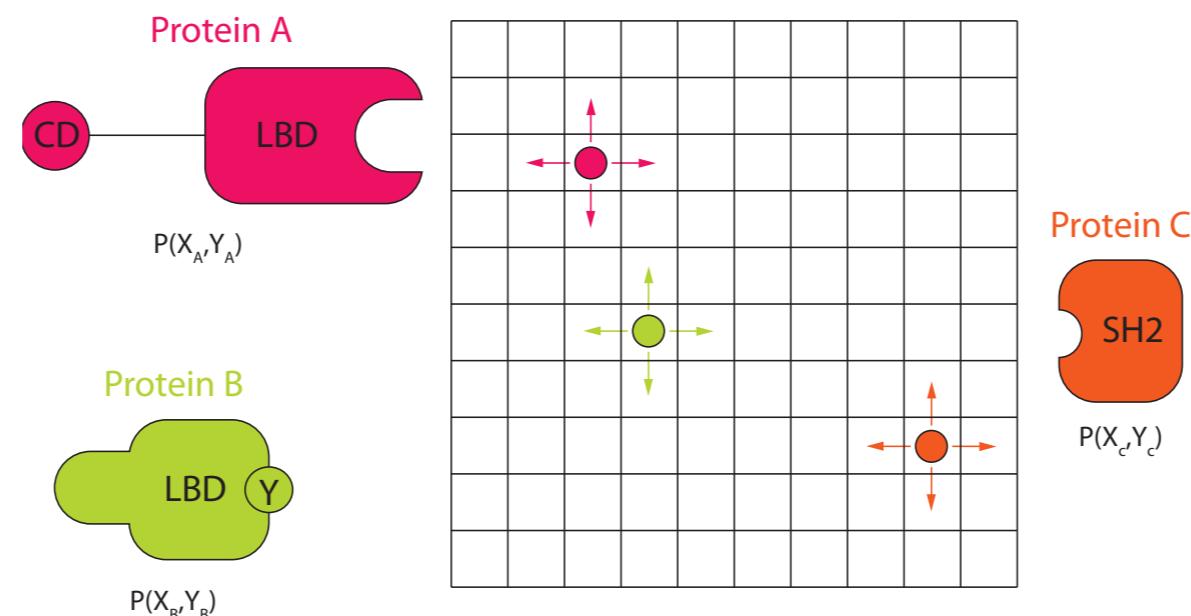
Restricted Interaction - Only if ProteinB (X,Y) = ProteinC (X,Y)



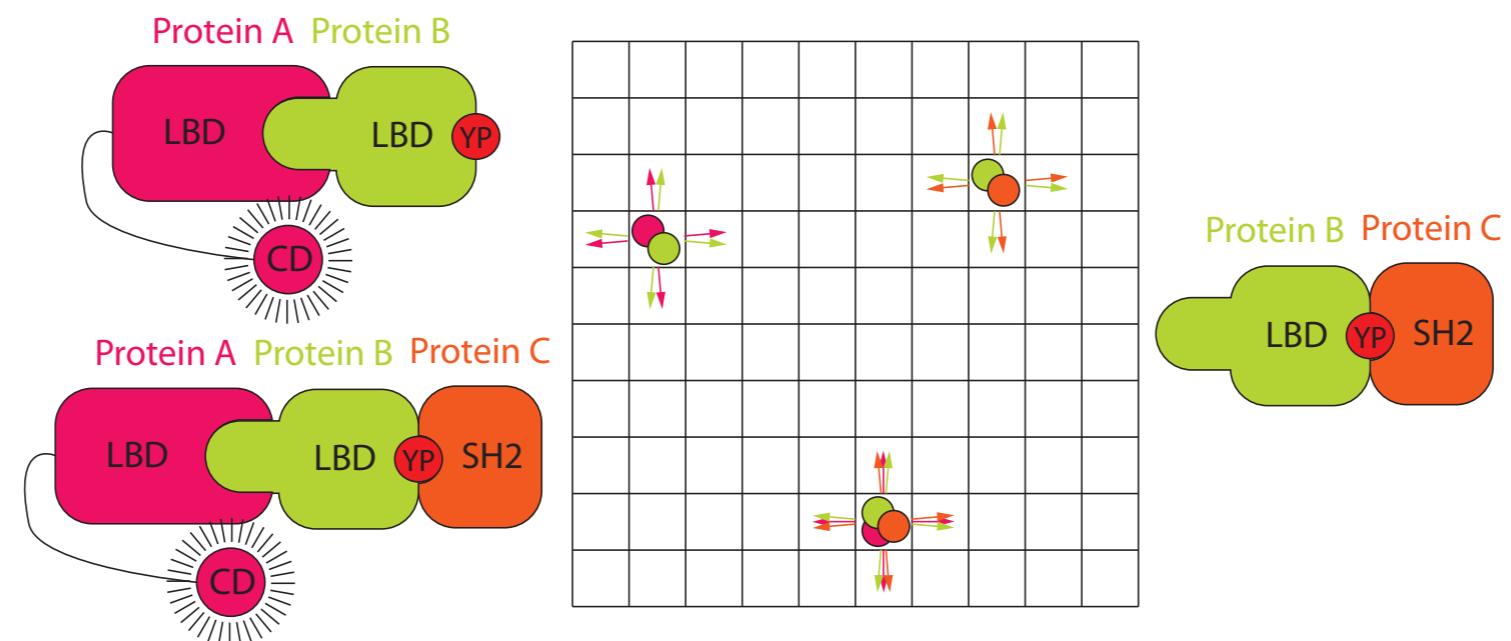
$$(ProteinB_X = ProteinC_X) \& (ProteinB_Y = ProteinC_Y)^* h(t)$$

Step 3: Explicit Encoding of Local Position Changes

Case 1: Movement of components as a single entity

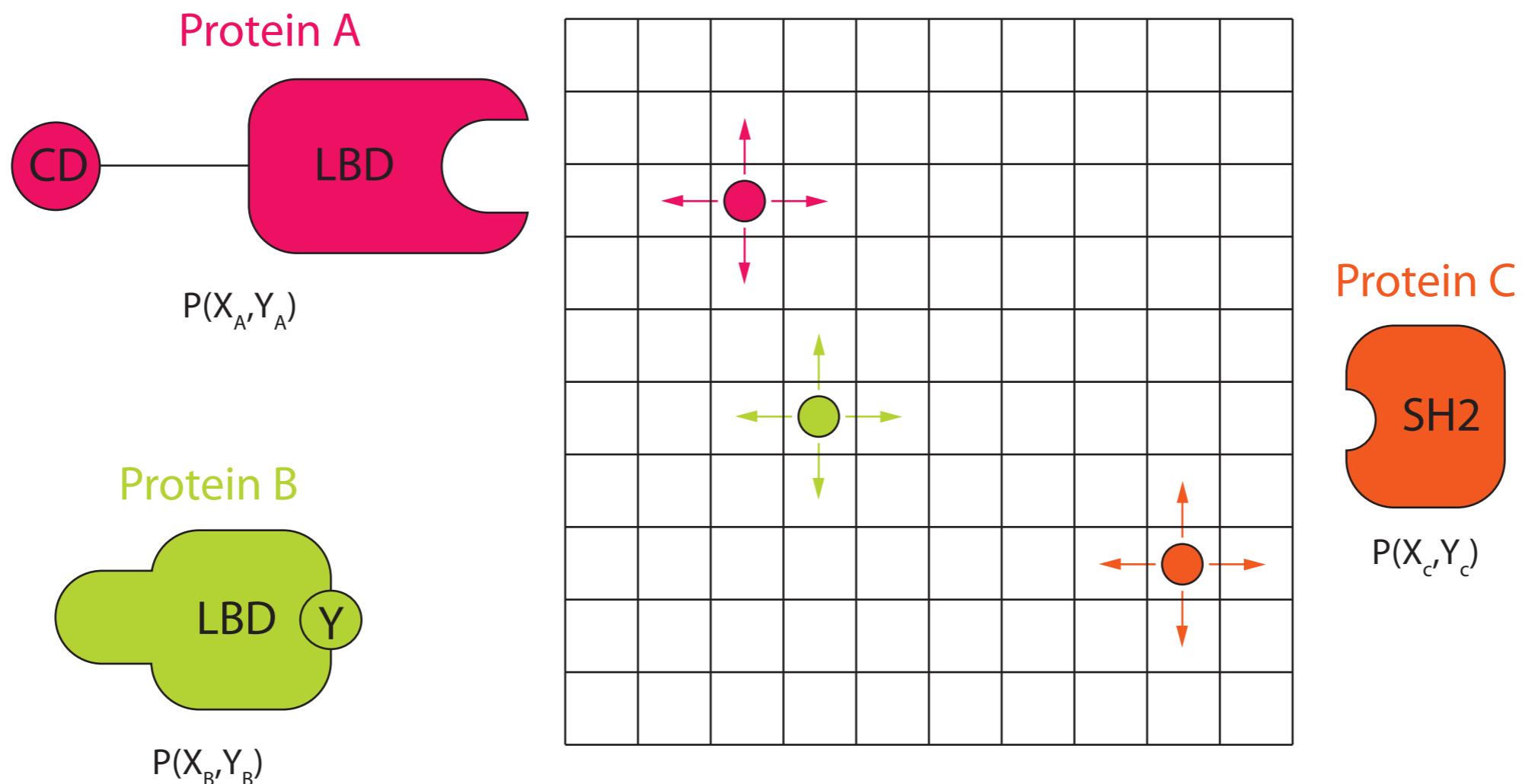


Case 2: Movement of components as complex



Case 1: Movement of components as a single entity

- Movement along the axes in respect to the defined grid size
 - e.g. 2D-Grid -> 2 direction of movement per axis
- Movement only if all interaction sites are unused
 - e.g. Protein B is not allowed to interact with Protein A or Protein C

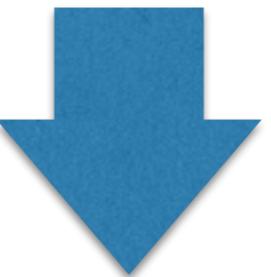


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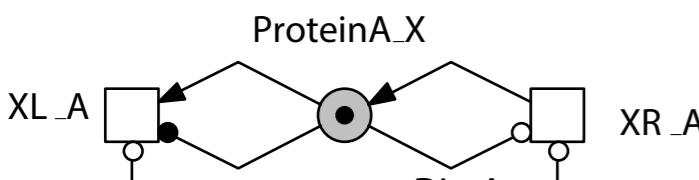


For each component:

- add two transitions per axis to increase/decrease the value of the coordinate places in respect to the grid size
- connect places representing interaction states of the components via inhibitory arcs

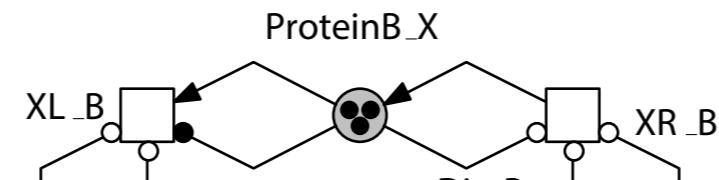
Case 1: Movement of components as a single entity

Movement of Protein A



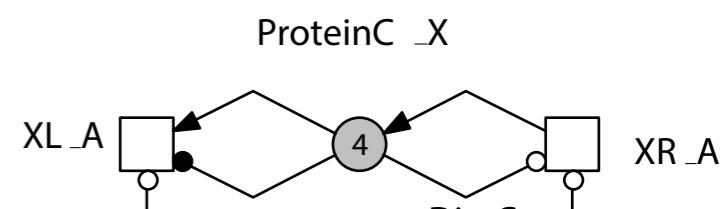
ProteinA_LBD .. ProteinB_LBD

Movement of Protein B

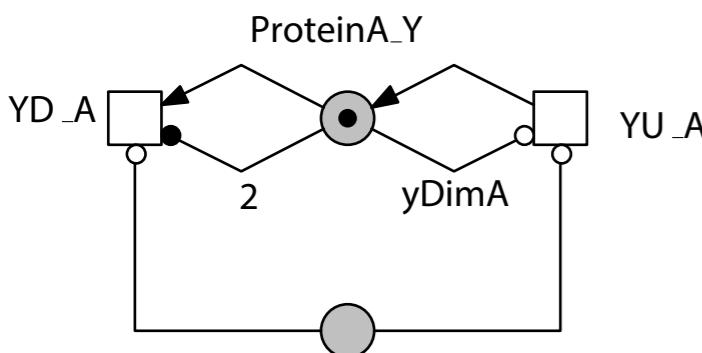


ProteinA_LBD .. ProteinB_LBD

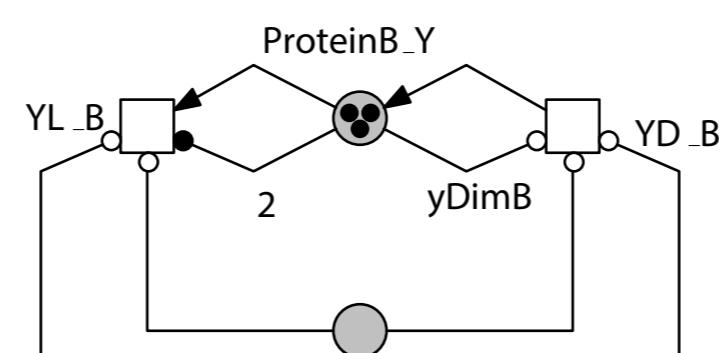
Movement of Protein C



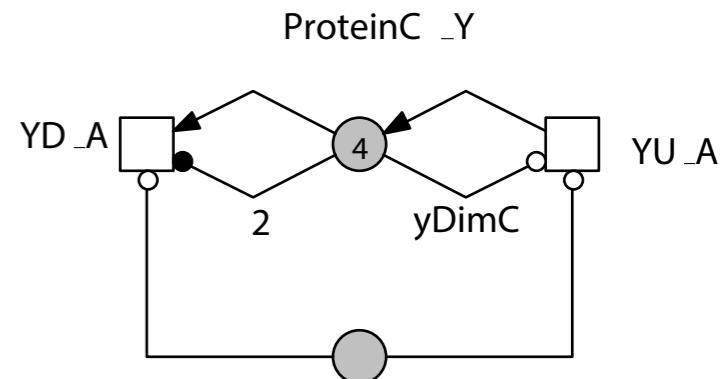
ProteinB_TYRp .. ProteinC_SH2



ProteinA_LBD .. ProteinB_LBD



ProteinA_LBD .. ProteinB_LBD

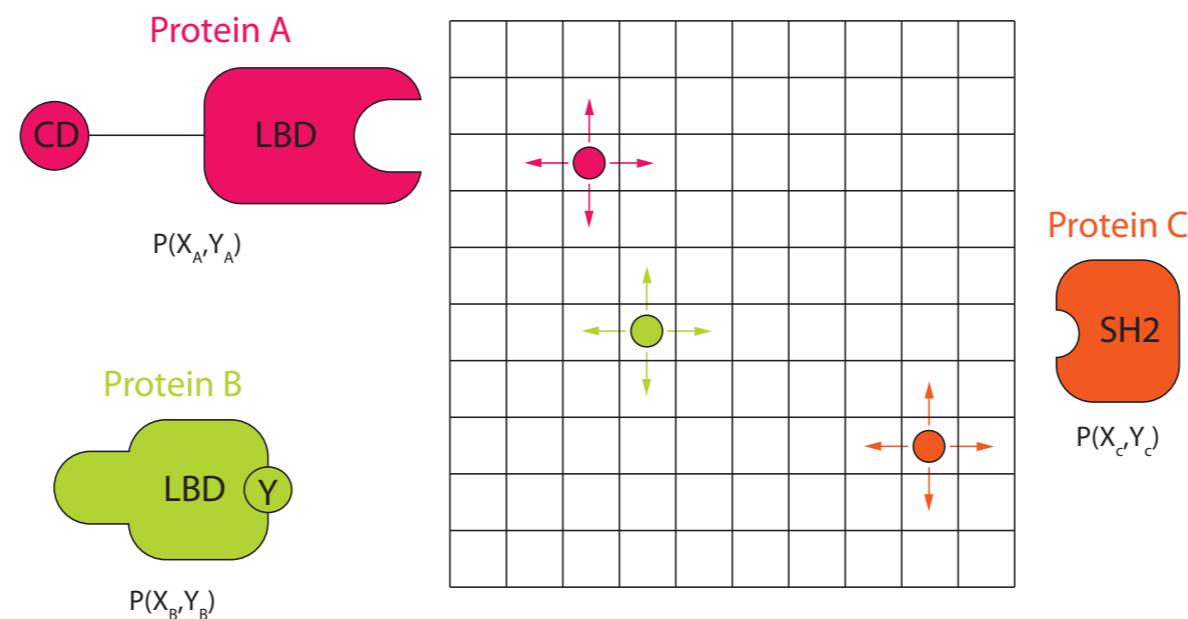


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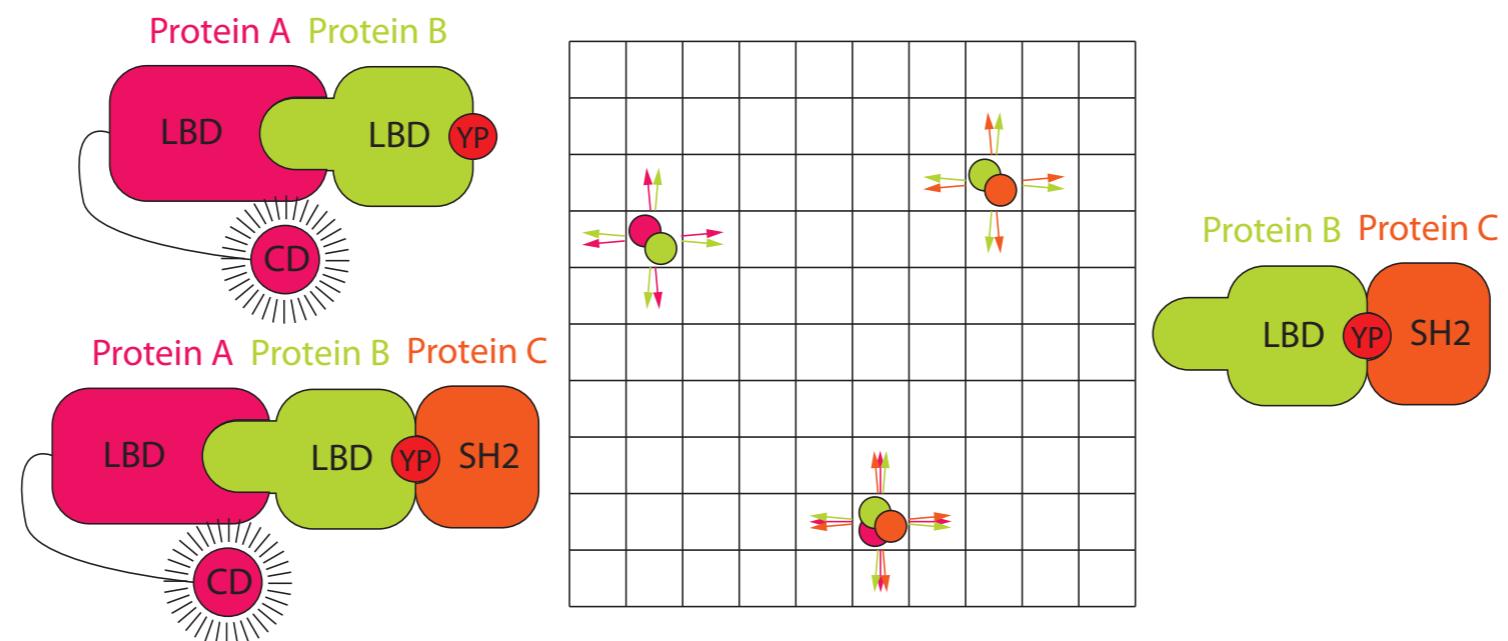
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Case 1: Movement of components as a single entity

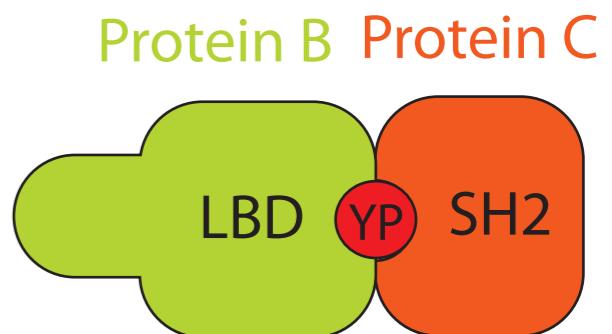
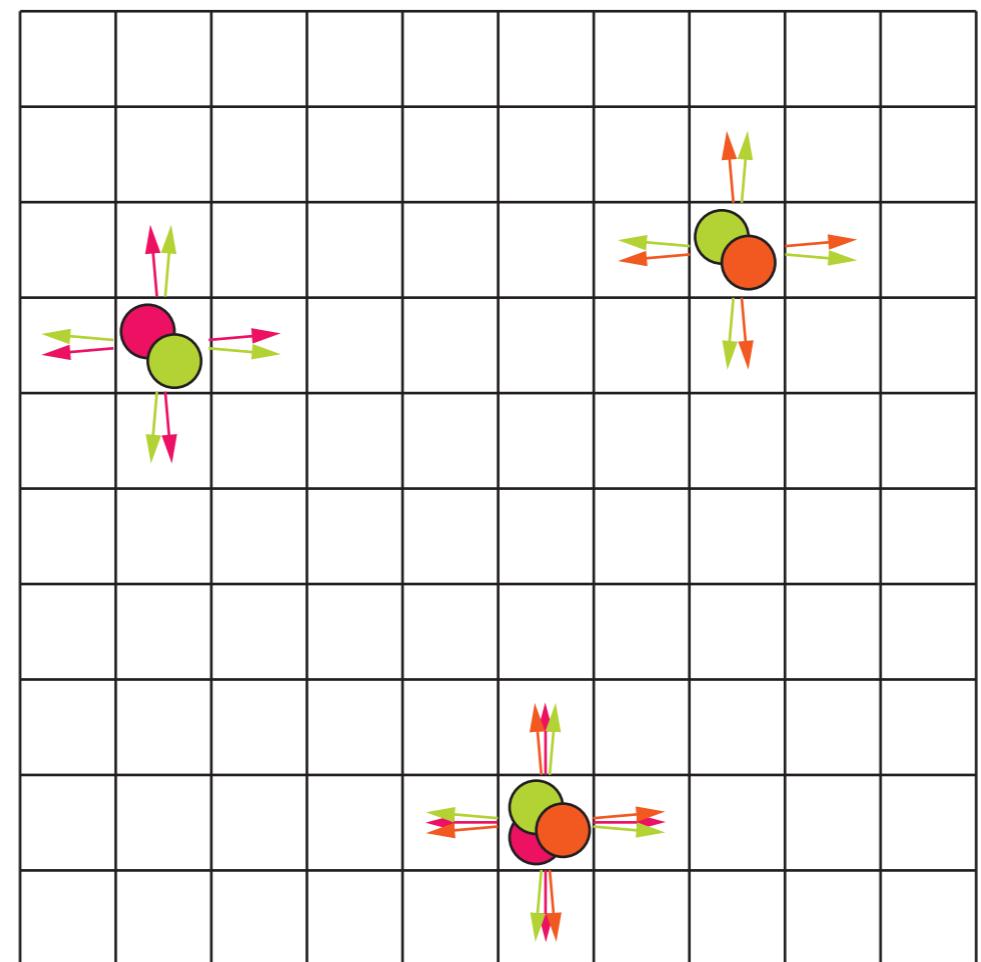
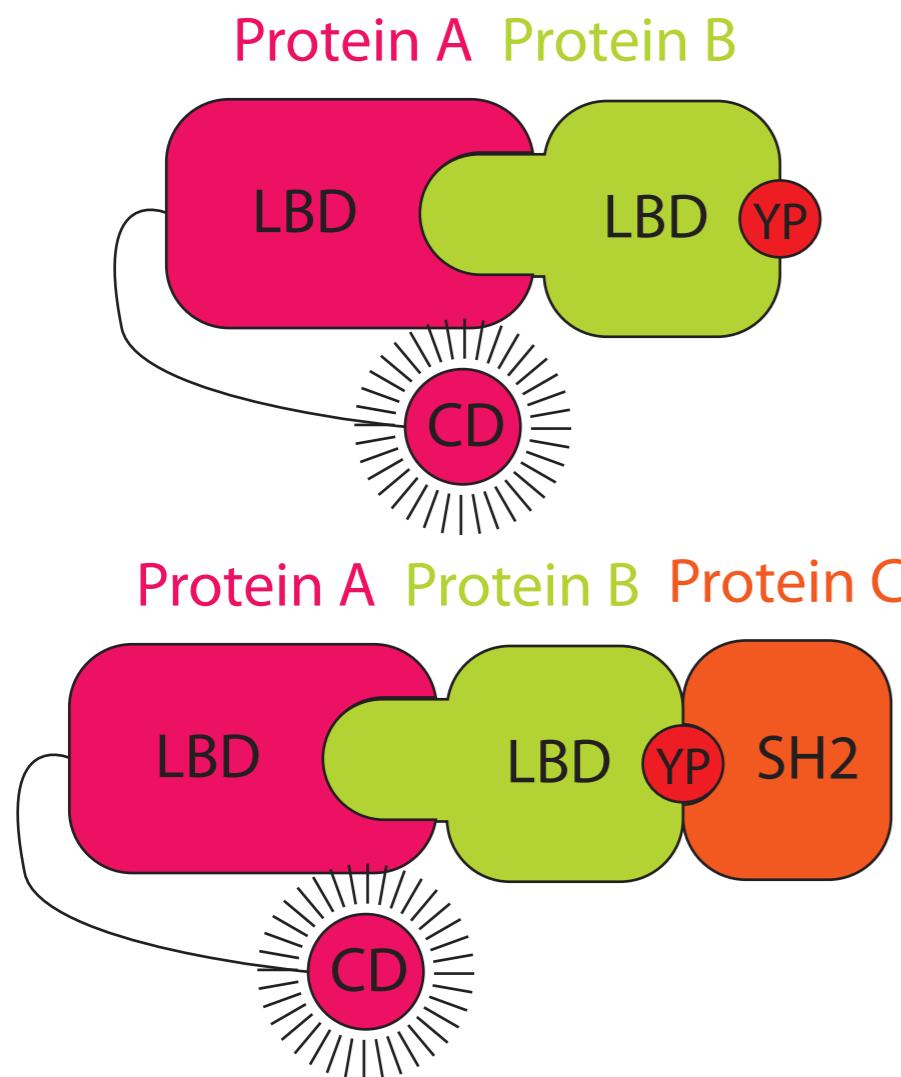


Case 2: Movement of components as complex



Case 2: Movement of components as complex

- Determine all possible complexes
- e.g. Protein A + Protein B, Protein B + Protein C, Protein A + Protein B + Protein C
- Movement only if the corresponding interaction sites of interacting components in a complex are used and all other are unused
- e.g. Protein A + Protein B can only move if they interact with each other, but not with Protein C
- Update local positions of all components forming a complex simultaneously

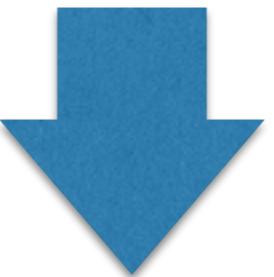


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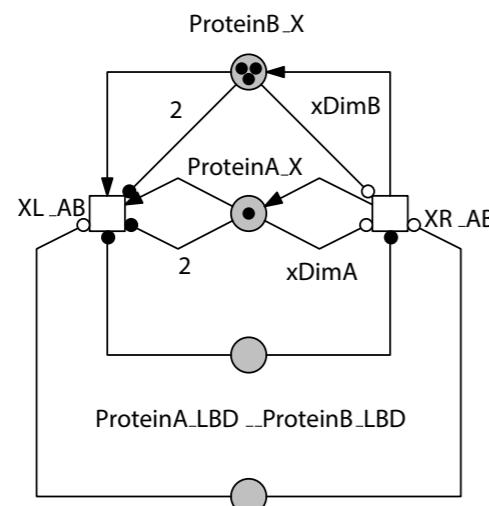


For each complex:

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- connect places representing the respective interaction via test arcs
- connect places representing interaction with other components via inhibitory arcs

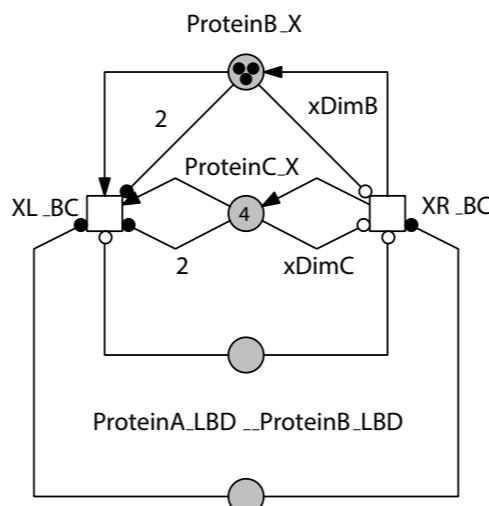
Case 2: Movement of components as complex

Movement of complex ProteinA_ProteinB



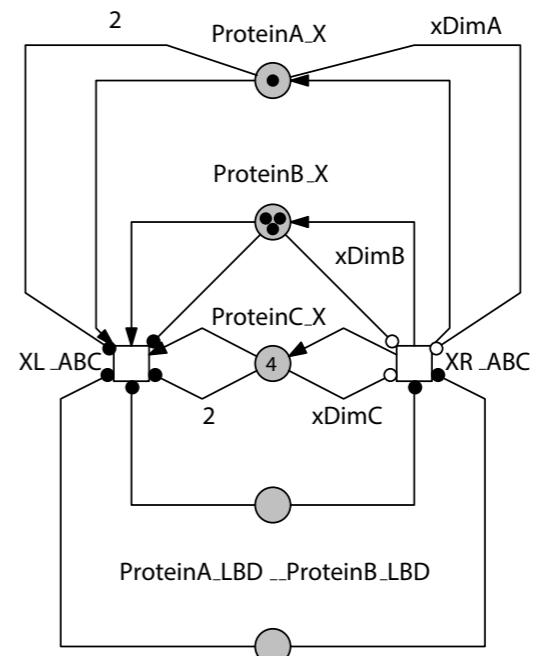
ProteinB_TYRp .. ProteinC_SH2

Movement of complex ProteinB_ProteinC

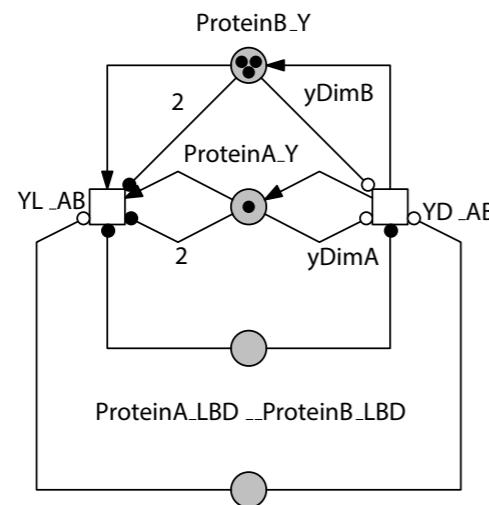


ProteinB_TYRp .. ProteinC_SH2

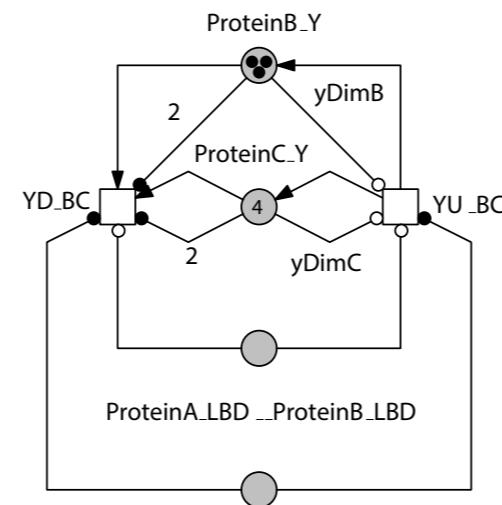
Movement of complex ProteinA_ProteinB_ProteinC



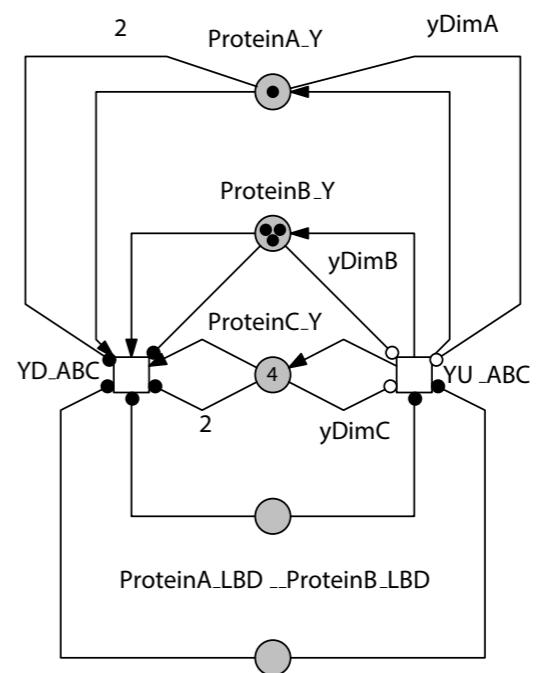
ProteinB_TYRp .. ProteinC_SH2



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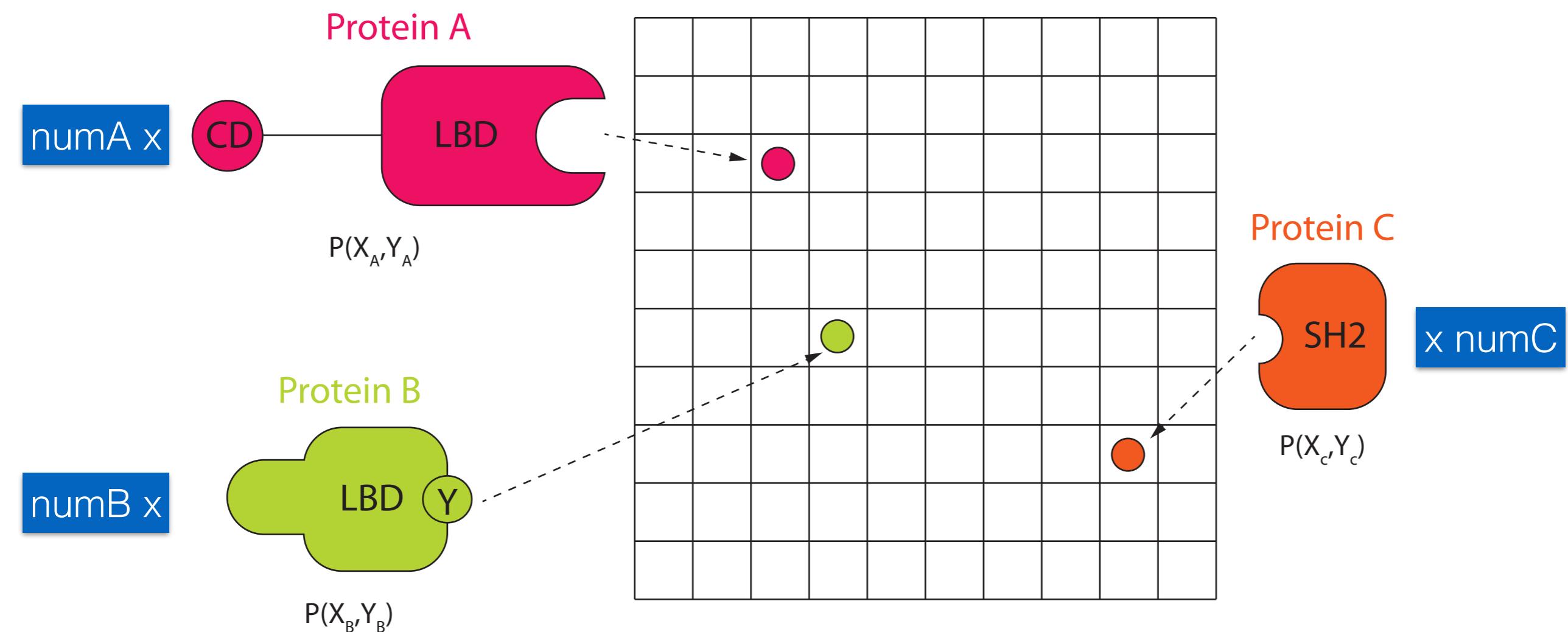
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ProteinB_TYRp .. ProteinC_SH2

Explicit Encoding of Component Instances

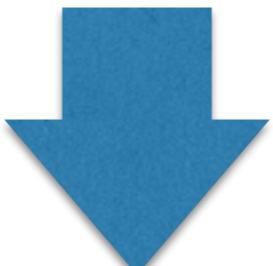
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 - e.g. numA = 3, numB= 3, numC = 3
- Duplicate the previously defined networks according to the number of instances



Explicit Encoding of Component Instances

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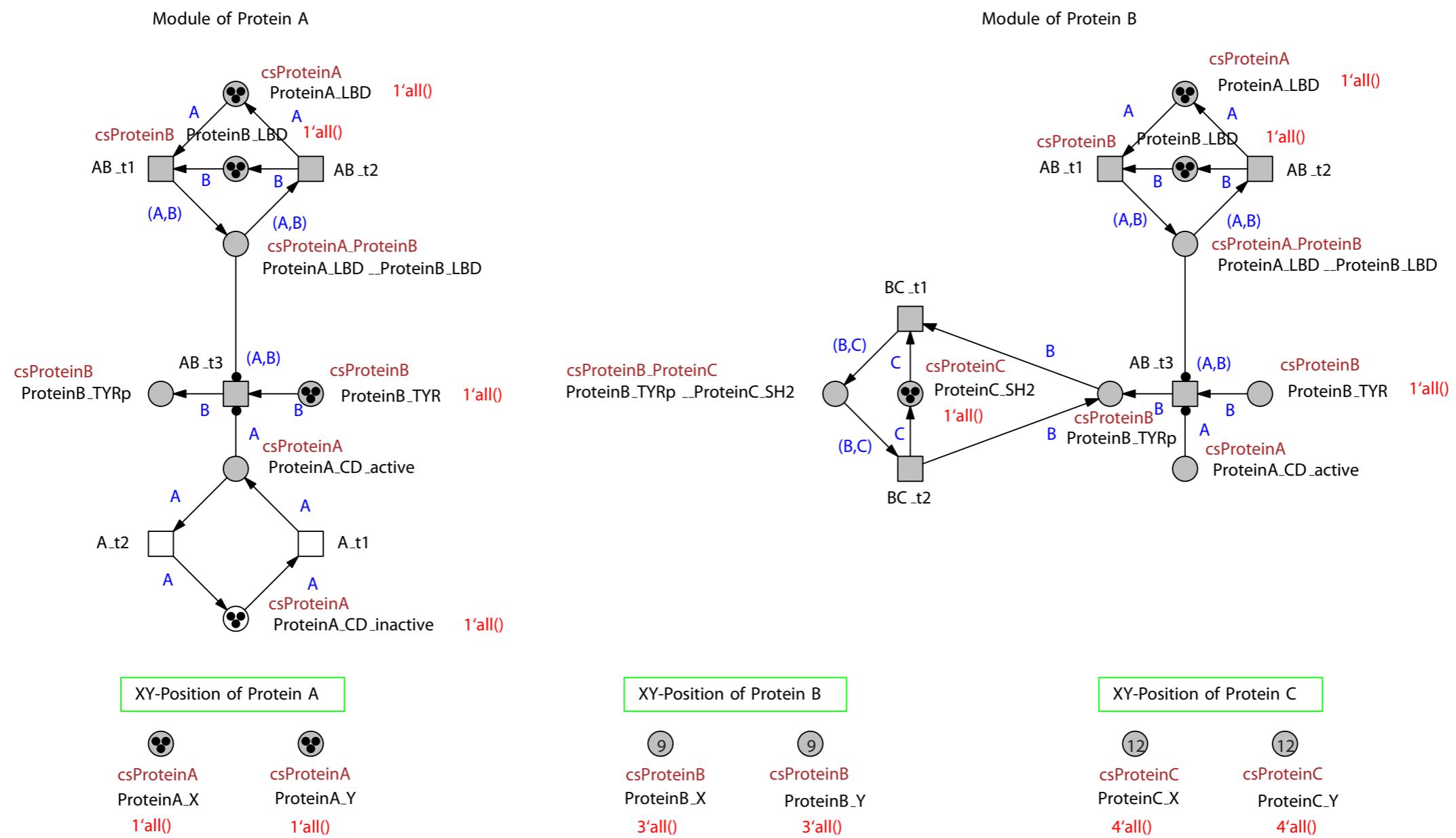
Explicit Encoding of Component Instances

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 - e.g. numA = 3, numB= 3, numC = 3
 - Duplicate the previously defined networks according to the number of instances
- 
- Application of coloured Petri nets:
- For each component define a simple colour-set and variable,
 - Add simple colour-sets to places representing non-interaction states
 - For each interaction define a product colour-set based on the respective simple colour-sets
 - Add product colour-sets to all places representing the respective interaction

Applying Coloured Petri Nets

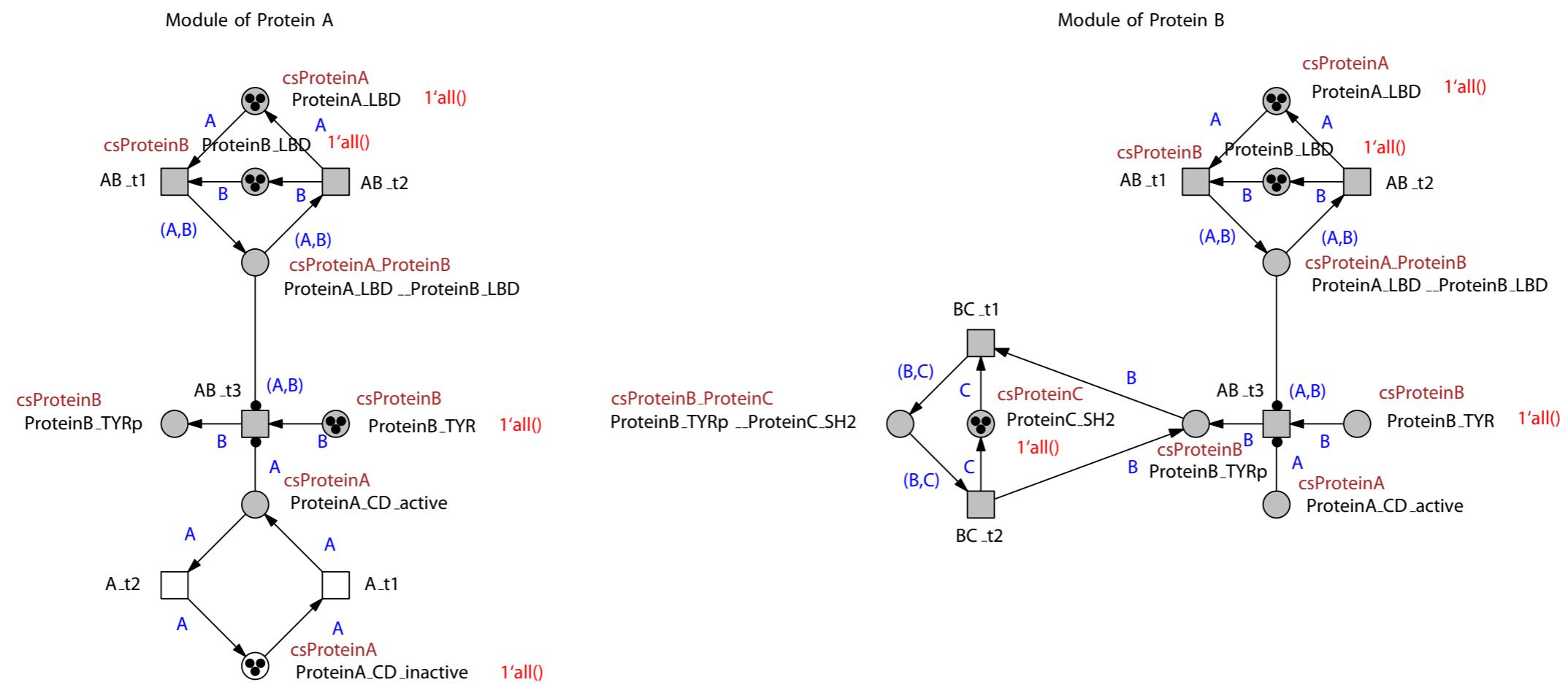
- Define a simple colour-set and a variable for each component

- Protein A** -> csProteinA:= int, 1-numA; csProteinA A
- Protein B** -> csProteinB:= int, 1-numB; csProteinB B
- Protein C** -> csProteinC:= int, 1-numC; csProteinC C



Applying Coloured Petri Nets

- Define a product colour-set for each interaction
- Protein A + Protein B \rightarrow csProteinA_ProteinB := product csProteinA, csProteinB
- Protein B + Protein C \rightarrow csProteinB_ProteinC := product csProteinB, csProteinC



XY-Position of Protein A

	csProteinA
ProteinA_X	
1'all()	

	csProteinA
ProteinA_Y	
1'all()	

XY-Position of Protein B

	csProteinB
ProteinB_X	
3'all()	

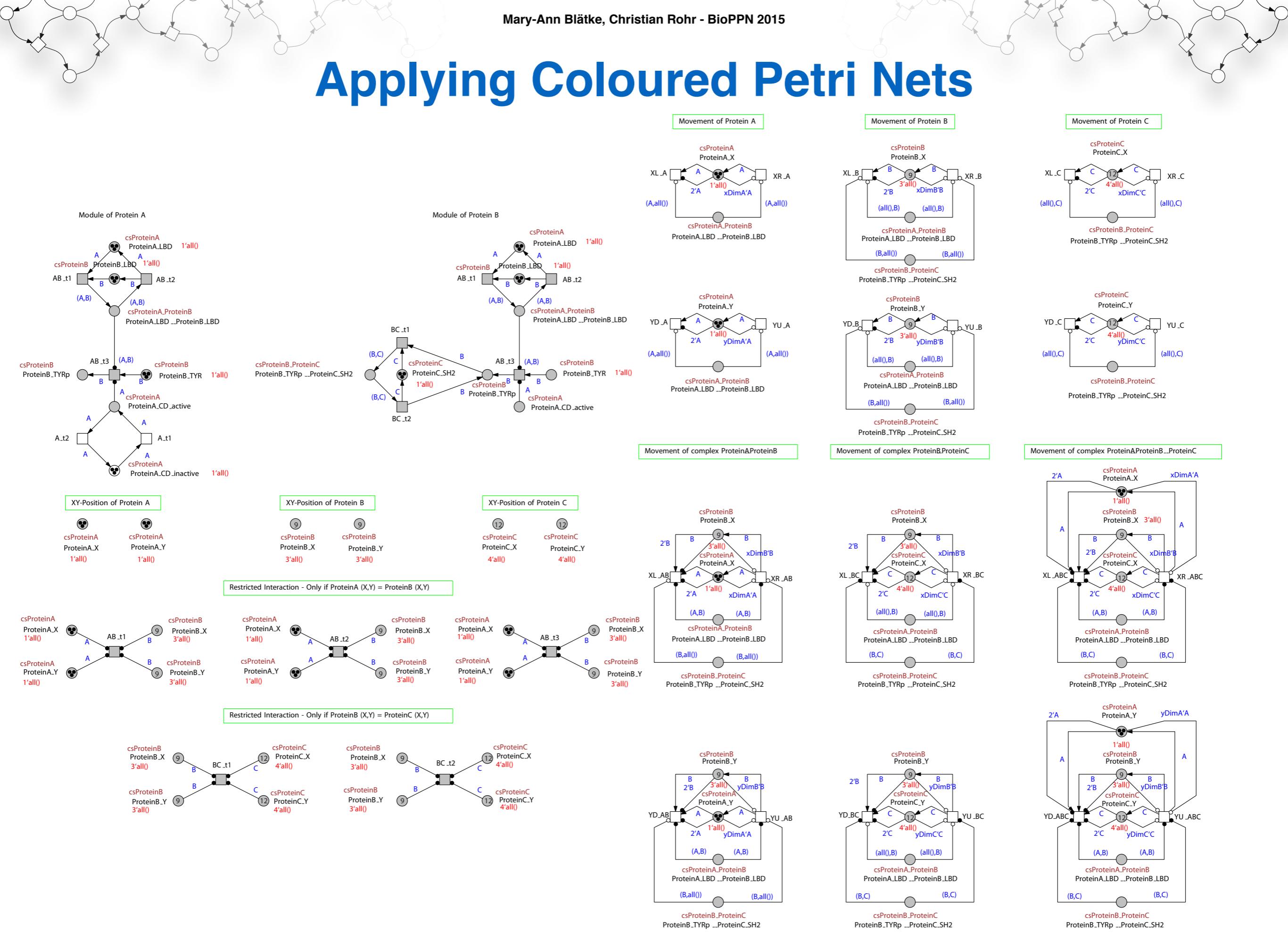
	csProteinB
ProteinB_Y	
3'all()	

XY-Position of Protein C

	csProteinC
ProteinC_X	
4'all()	

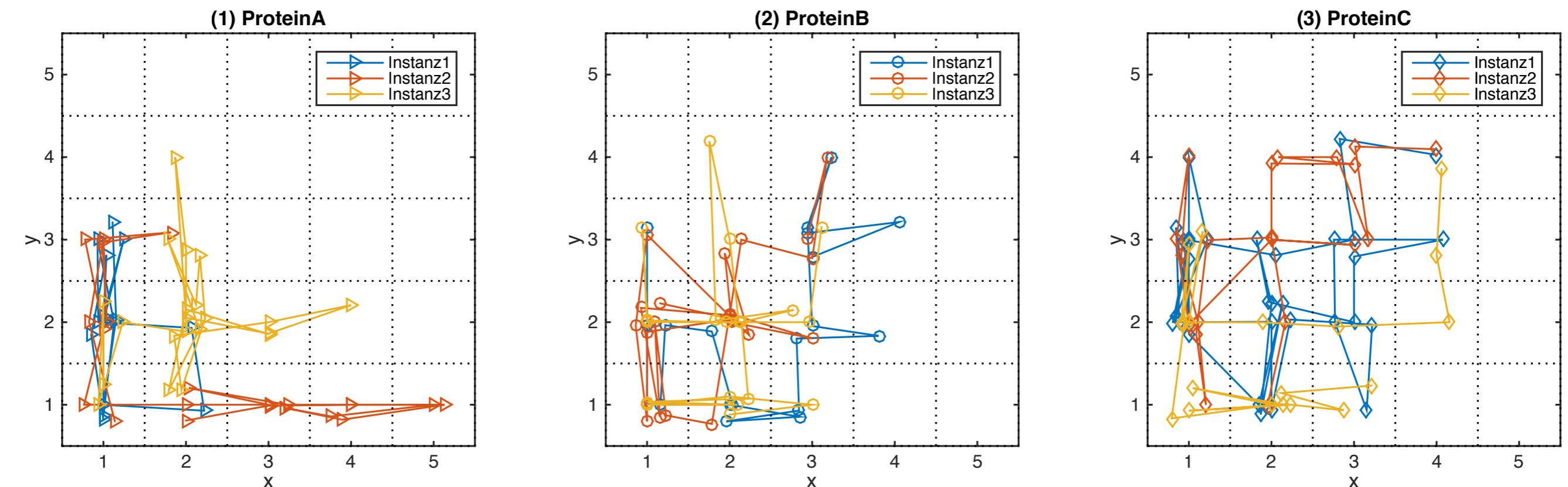
	csProteinC
ProteinC_Y	
4'all()	

Applying Coloured Petri Nets

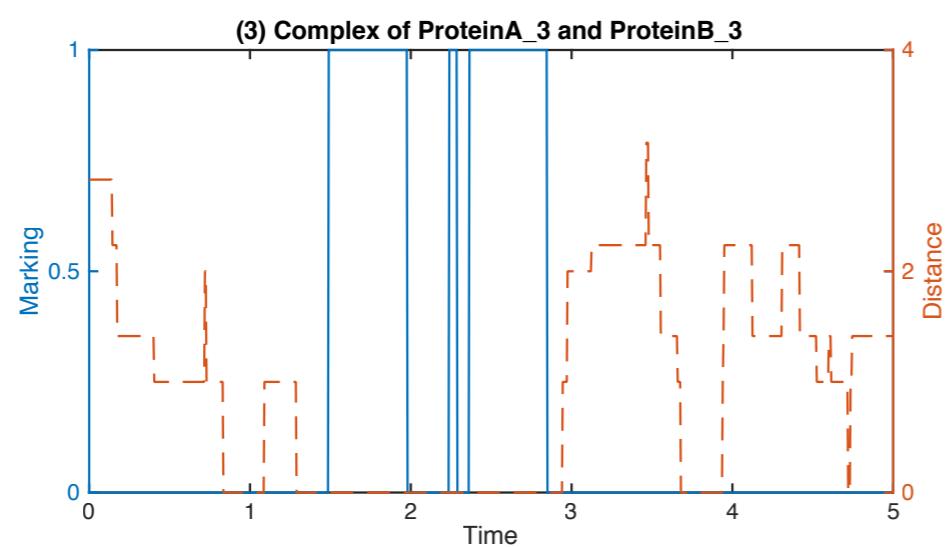
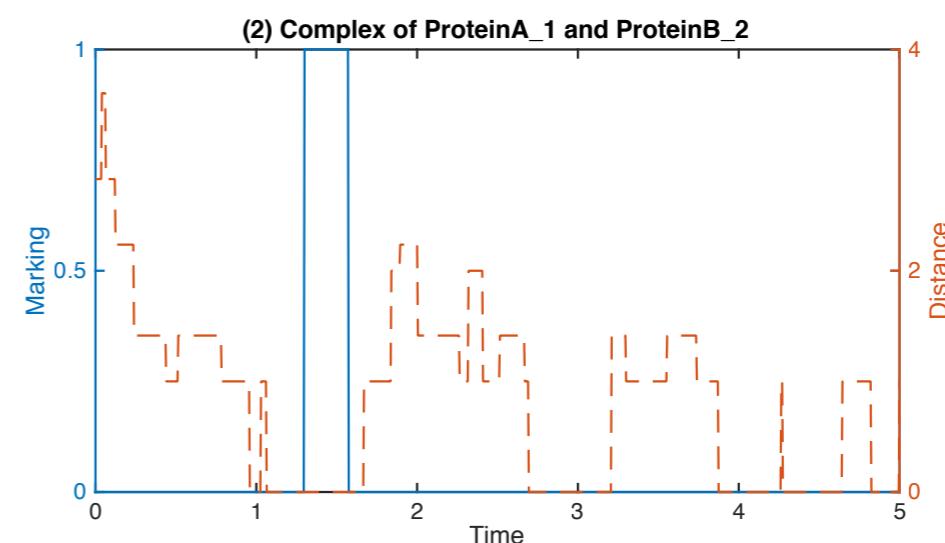
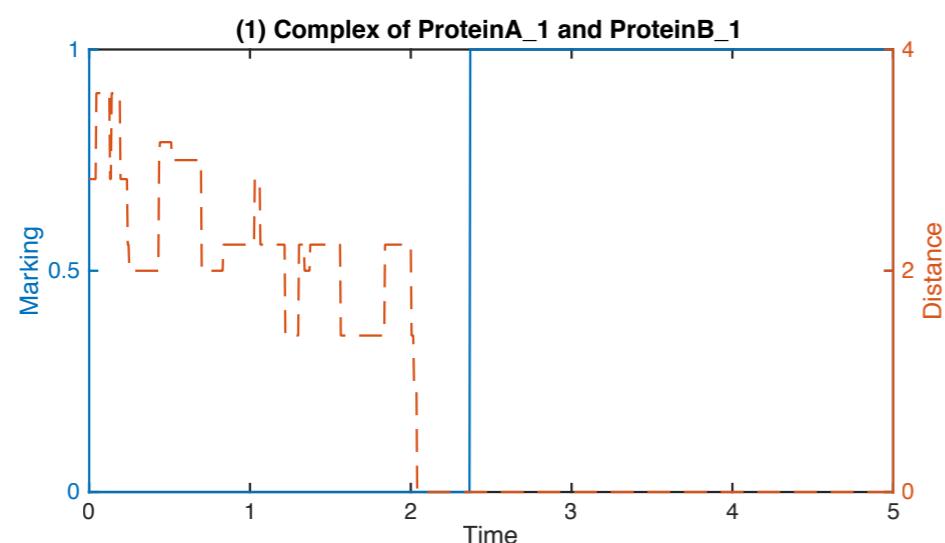


Stochastic Simulation

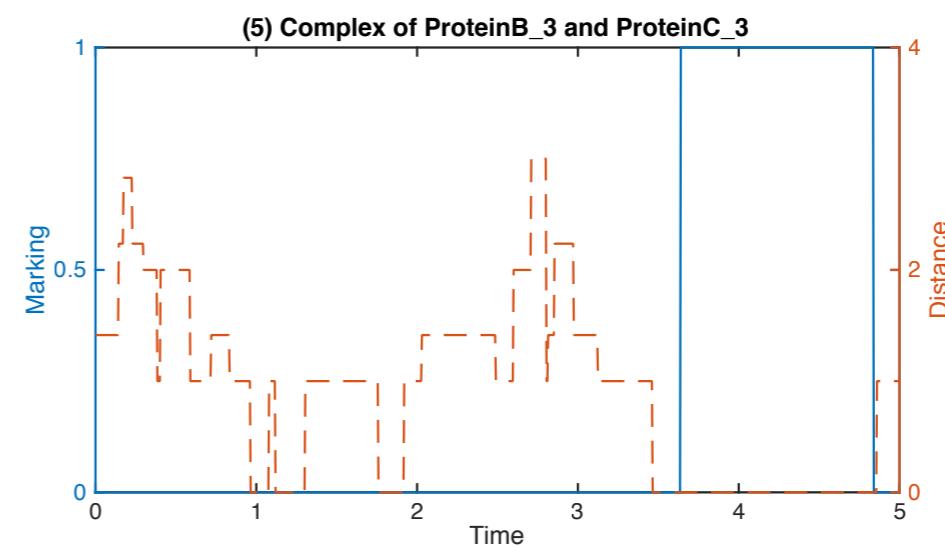
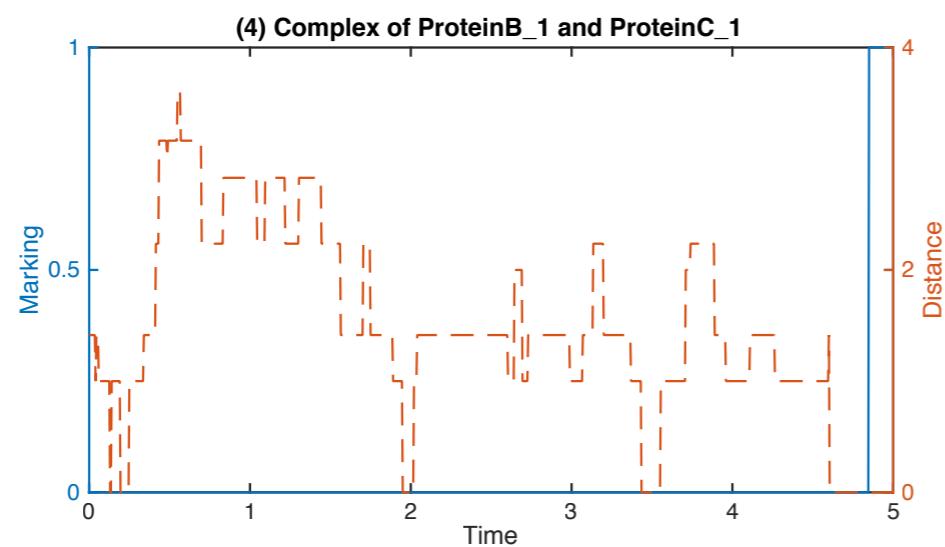
- Model parameter:
 - 5x5 2D-Grid for each component
 - 3 instances per component
- Model size:
 - $|P|=57$, $|T|=267$, $|A|=1992$



Stochastic Simulation



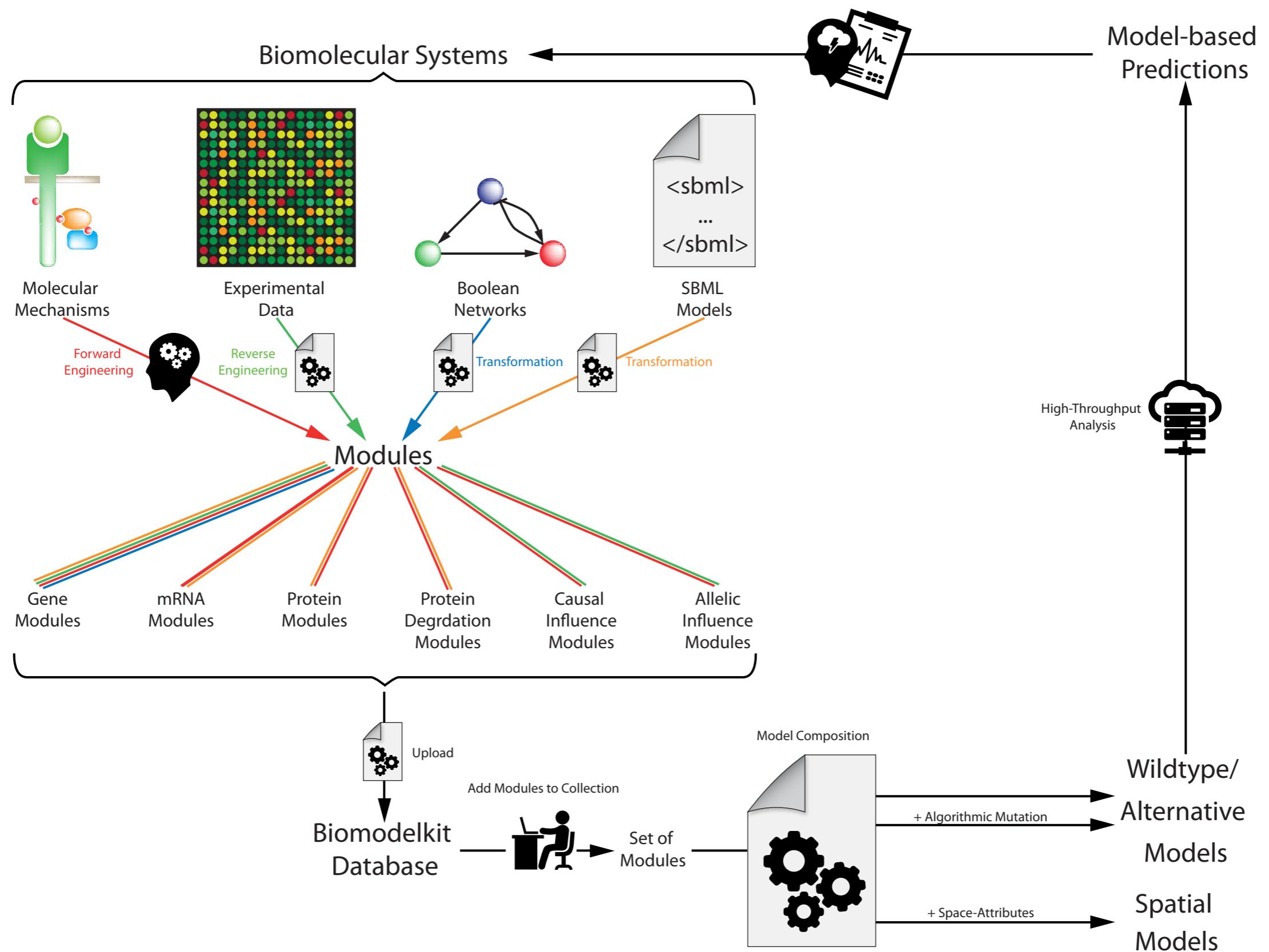
Components move as one entity while interacting



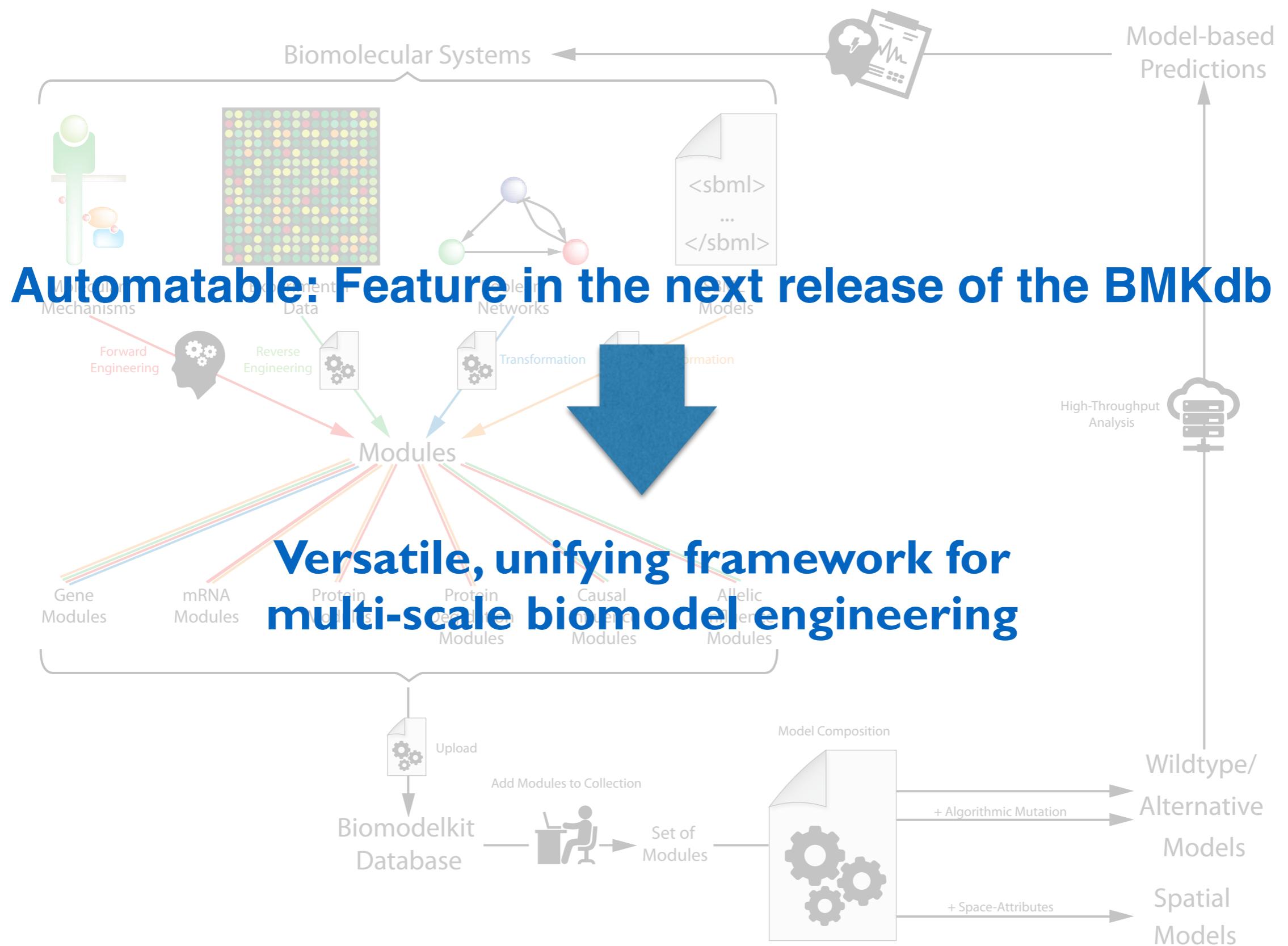
Summary

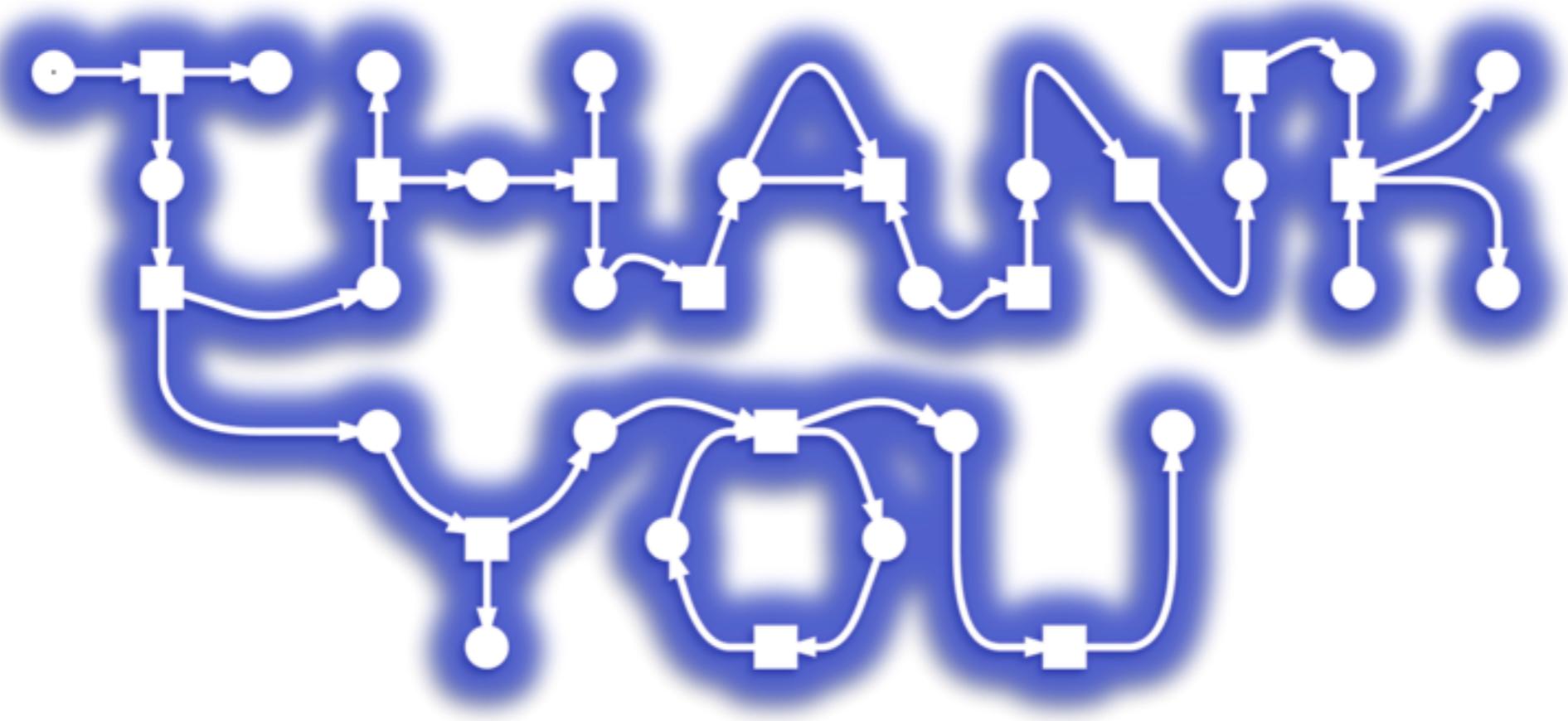
- Spatial Transformation Algorithm
 1. Explicit encoding of local positions
 2. Local restriction of interactions
 3. Explicit encoding of local position changes
 4. Explicit encoding of component instances using coloured Petri nets
- No interference with the model structure
 - Transformation is reversible
- Not restricted to discrete space
 - Representation of continuous space by the use of continuous places

Summary



Summary





Cooperation Partners

Monika Heiner and Co-Workers, BTU Cottbus
David Gilbert, Brunel University London
Fred Scharper and Co-Workers, OvGU Magdeburg
Tim Hucho, University of Cologne
Fei Liu, Harbin Institute of Technology

Projects

Consortium „Modelling of Pain Switches“ 2009-2011
Consortium „NoPain“ 2013-2015

Graduate School

IMPRS Magdeburg



Sachsen-Anhalt
Centre for Systems Biology
MaCS



Federal Ministry
of Education
and Research

FORSYS
Research Units for Systems Biology



DFG

Deutsche
Forschungsgemeinschaft

IMPRS ProEng | 
International Max Planck Research School for
Advanced Methods in Process and Systems Engineering
Magdeburg | 

Recent Publications



Chapter 6
A Petri-Net-Based Framework for Biomodel Engineering

Mary Ann Blätke, Christian Rohr, Monika Heiner, and Wolfgang Marwan

Abstract Petri nets provide a unifying and versatile framework for the synthesis and engineering of computational models of biochemical reaction networks and of gene regulatory networks. Starting with the basic definitions, we provide an introduction into the different classes of Petri nets that reinterpret a Petri net graph as a qualitative, stochastic, continuous, or hybrid model. Static and dynamic analysis in addition to simulative model checking provide a rich choice of methods for the analysis of the structure and dynamic behavior of Petri net models. Coloring of Petri nets of all classes is powerful for multiscale modeling and for the representation of location and space in reaction networks since it combines the concept of Petri nets with the computational mightiness of a programming language. In the context of the Petri net framework, we provide two most recently developed approaches to biomodel engineering, the database-assisted automatic composition and modification of Petri nets with the help of reusable, automatic reconstruction of networks based features the framework provides multiple options for the context of systems and synthetic biology.

Keywords Automatic network reconstruction · Systems modelling · Modular modelling · Petri networks · Reverse engineering

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Birkhäuser

Chapter 7



BioModel Engineering with Petri Nets

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7.1 INTRODUCTION

BioModel Engineering

Systematically constructing, maintaining, and deploying artifacts are typical attributes of sound engineering, independent of the application field. Along these lines, BioModel Engineering is the science of engineering computational models of biochemical processes in an efficient, sustainable, and trustworthy manner.

In *Systems Biology*, models are used to describe our abstract understanding of biochemical processes and to predict their behavior, for example, in response to perturbations like mutations, chemical interventions, or changes in the environment. In *Synthetic Biology*, models support the reliable design and redesign of molecular networks and may serve as design templates for models have high explanatory and predictive power.

In this chapter, we will explore how this aim the unifying power of Petri nets, serving as umbrella continuous, and hybrid paradigms. An introductory transduction pathways can be found in [1]; a more rigorous treatment is given in [2].

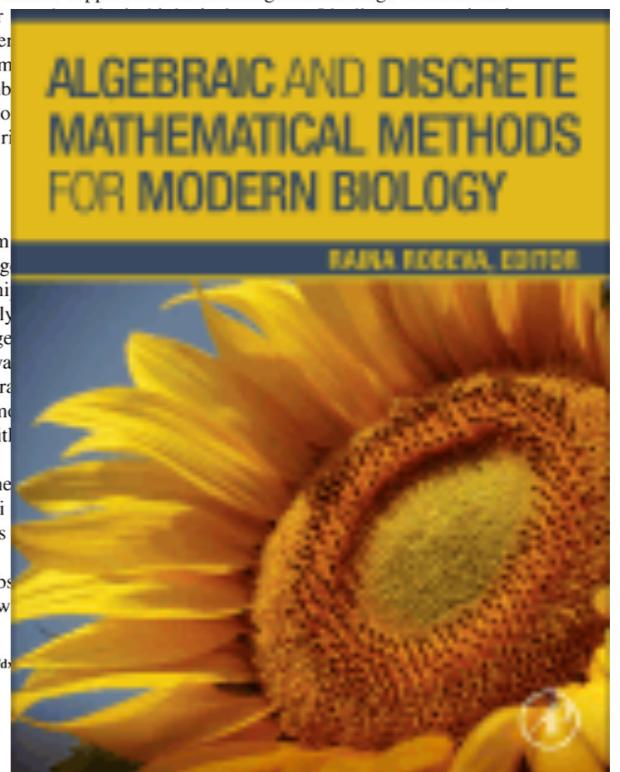
Petri Nets

The basic ideas of Petri nets, as we understand them in 1962 [3]. Petri nets are basically a formal language semantics. They permit the modeling of relationships between transitions in the Petri net terminology and typically interpreted as any (bio-)chemical species, such as genes, whereas events model (bio-)chemical reactions at various levels of abstraction. Petri nets support regulatory activation and deactivation, transport, transitions, stoichiometry of (bio-)chemical reactions or the amorphous structure of a Petri net is unambiguous and explicitly concurrency.

Enhancing these key modeling principles by the from programming languages, yields colored Petri “discrete data type.” They are particularly strong, as similar processes in similar components.

Petri nets may represent species on different levels of abstraction, from unicellular organisms to multicellular organisms and populations. This allows the

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Scaling Properties

- The unfolded model scales with:
 - number of axes (1D, 2D, 3D)
 - number of instances and the resulting number of complexes among the components

Number of Instances	$n = 1$	$n = 5$	$n = 10$	$n = 50$	$n = 100$
Places	15	115	330	5650	21300
Transitions	31	895	5440	533200	4131400
Arcs	160	6880	42760	4253800	33007600
Unfolding Time (s)	0.124	0.305	1.536	180.763	1608.802